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### (54) Thermostable DNA polymerases

(57) An enzymatically active DNA polymerase having between 540 and 582 amino acids having a tyrosine at a position equivalent to position 667 of Taq DNA polymerase, wherein said polymerase lacks 5' to 3' exonuclease activity, and wherein said polymerase has at

least 95% homology in its amino acid sequence to the DNA polymerase of Thermus aquaticus, Thermus flavus or Thermus thermophilus, and wherein said polymerase forms a single polypeptide band on an SDS PAGE.

**Description**Background of the Invention

5 The present invention relates to novel thermostable DNA polymerases, the genes and vectors encoding them and their use in DNA sequencing.

10 US Patents 4,889,818 and 5,079,352 describe the isolation and expression of a DNA polymerase known as Taq DNA Polymerase (hereinafter referred to as Taq). It is reported that amino-terminal deletions wherein approximately one-third of the coding sequence is absent have resulted in producing a gene product that is quite active in polymerase assays. Taq is described as being of use in PCR (polymerase chain reaction).

15 US Patent No. 5,075,216 describes the use of Taq in DNA sequencing.

20 International patent application WO 92/06/06188 describes a DNA polymerase having an identical amino acid sequence to Taq except that it lacks the N-terminal 235 amino acids of Taq and its use in sequencing. This DNA polymerase is known as  $\Delta$  Taq.

25 US Patent 4,795,699 describes the use of T7 type DNA polymerases (T7) in DNA sequencing. These are of great use in DNA sequencing in that they incorporate dideoxy nucleoside triphosphates (NTPs) with an efficiency comparable to the incorporation of deoxy NTPs; other polymerases incorporate dideoxy NTPs far less efficiently which requires comparatively large quantities of these to be present in sequencing reactions.

30 At the DOE Contractor-Grantee Workshop (Nov. 13-17, 1994, Santa Fe) and the I. Robert Lehman Symposium (Nov 11-14, 1994, Sonoma), Prof. S. Tabor identified a site in DNA polymerases that can be modified to incorporate dideoxy NTPs more efficiently. He reported that the presence or absence of a single hydroxy group (tyrosine vs. phenylalanine) at a highly conserved position on *E. coli*, DNA Polymerase I, T7, and Taq makes more than a 1000-fold difference in their ability to discriminate against dideoxy NTPs. (See also European Patent Application 94203433.1 published May 31, 1995, Publication No. 0 655 506 A1 and hereby incorporated by reference herein.)

Summary of the Invention

35 The present invention provides a DNA polymerase having an amino acid sequence differentiated from Taq in that it lacks the N-terminal 272 amino acids and has the phenylalanine at position 667 (of native Taq) replaced by tyrosine. Preferably, the DNA polymerase has methionine at position 1 (equivalent to position 272 of Taq) (hereinafter referred to as FY2) The full DNA sequence is given as Fig 1 (SEQ. ID. NO. 1). Included within the scope of the present invention are DNA polymerases having substantially identical amino acid sequences to the above which retain thermostability and efficient incorporation of dideoxy NTPs.

40 By a substantially identical amino acid sequence is meant a sequence which contains 540 to 582 amino acids that may have conservative amino acid changes compared with Taq which do not significantly influence thermostability or nucleotide incorporation, i.e. other than the phenylalanine to tyrosine conversion. Such changes include substitution of like charged amino acids for one another, or amino acids with small side chains for other small side chains, e.g., ala for val. More drastic changes may be introduced at noncritical regions where little or no effect on polymerase activity is observed by such a change.

45 The invention also features DNA polymerases that lack between 251 and 293 (preferably 271 or 272) of the N-terminal amino acids of *Thermus flavus* (Tfl) and have the phenylalanine at position 666 (of native Tfl) replaced by tyrosine; and those that lack between 253 and 295 (preferably 274) of the N-terminal amino acids of *Thermus thermophilus* (Tth) and have the phenylalanine at position 669 (of native Tth) replaced by tyrosine.

50 By efficient incorporation of dideoxy NTPs is meant the ability of a polymerase to show little, if any, discrimination in the incorporation of ddNTPs when compared with dNTPs. Suitably efficient discrimination is less than 1:10 and preferably less than 1:5. Such discrimination can be measured by procedures known in the art.

55 One preferred substantially identical amino acid sequence to that given above is that which contains 562 amino acids having methionine at position 1 and alanine at position 2 (corresponding to positions 271 and 272 of native Taq) (hereinafter referred to as FY3). A full DNA sequence is given as Fig. 2. This is a preferred DNA polymerase for expression by a gene of the present invention.

The purified DNA polymerases FY2 and FY3 both give a single polypeptide band on SDS polyacrylamide gels, unlike  $\Delta$  Taq, having either a phenylalanine or tyrosine at position 667 which forms several polypeptide bands of similar size on SDS polyacrylamide gels.

A second preferred substantially identical amino acid sequence is that which lacks 274 of the N-terminal amino acids of *Thermus thermophilus* having methionine at position 1 and the phenylalanine to tyrosine mutation at position 396 (corresponding to position 669 of native Tth) (hereinafter referred to as FY4). A full DNA sequence is given as Fig. 5 (SEQ. ID. NO. 14).

The present invention also provides a gene encoding a DNA polymerase of the present invention. In order to assist

in the expression of the DNA polymerase activity, the modified gene preferably codes for a methionine residue at position 1 of the new DNA polymerase. In addition, in one preferred embodiment of the invention, the modified gene also codes for an alanine at position 2 (corresponding to position 272 of native Taq).

In a further aspect, the present invention provides a vector containing the gene encoding the DNA polymerase activity of the present invention, e.g., encoding an amino acid sequence differentiated from native Taq in that it lacks the N-terminal 272 amino acids and has phenylalanine at position 396 (equivalent to position 667 of Taq) replaced by tyrosine or a substantially identical amino acid sequence thereto.

In a yet further aspect, the present invention provides a host cell comprising a vector containing the gene encoding the DNA polymerase activity of the present invention, e.g., encoding an amino acid sequence differentiated from native Taq in that it lacks the N-terminal 272 amino acids and has phenylalanine at position 396 (equivalent to position 667 of native Taq) replaced by tyrosine or a substantially identical amino acid sequence thereto.

The DNA polymerases of the present invention are preferably in a purified form. By purified form is meant that the DNA polymerase is isolated from a majority of host cell proteins normally associated with it; preferably the polymerase is at least 10% (w/w) of the protein of a preparation, even more preferably it is provided as a homogeneous preparation, e.g., a homogeneous solution. Preferably the DNA polymerase is a single polypeptide on an SDS polyacrylamide gel.

The DNA polymerases of the present invention are suitably used in sequencing, preferably in combination with a pyrophosphatase. Accordingly, the present invention provides a composition which comprises a DNA polymerase of the present invention in combination with a pyrophosphatase, preferably a thermostable pyrophosphatase such as *Thermoplasma acidophilum* pyrophosphatase. (Schafer, G. and Richter, O.H. (1992) *Eur. J. Biochem.* **209**, 351-355).

The DNA polymerases of the present invention can be constructed using standard techniques. By way of example, mutagenic PCR primers can be designed to incorporate the desired Phe to Tyr amino acid change (FY mutation) in one primer. In our hands these primers also carried restriction sites that are found internally in the sequence of the Taq polymerase gene clone of Delta Taq, pWB253, which was used by us as template DNA. However, the same PCR product can be generated with this primer pair from any clone of Taq or with genomic DNA isolated directly from *Thermus aquaticus*. The PCR product encoding only part of the gene is then digested with the appropriate restriction enzymes and used as a replacement sequence for the clone of Delta Taq digested with the same restriction enzymes. In our hands the resulting plasmid was designated pWB253Y. The presence of the mutation can be verified by DNA sequencing of the amplified region of the gene.

Further primers can be prepared that encode for a methionine residue at the N-terminus that is not found at the corresponding position of Taq, the sequence continuing with amino acid residue 273. These primers can be used with a suitable plasmid, e.g., pWB253Y.DNA, as a template for amplification and the amplified gene inserted into a vector, e.g., pRE2, to create a gene, e.g., pRE273Y, encoding the polymerase (FY2). The entire gene can be verified by DNA sequencing.

Improved expression of the DNA polymerases of the present invention in the pRE2 expression vector was obtained by creating further genes, pREFY2pref (encoding a protein identical to FY2) and pREFY3 encoding FY3. A mutagenic PCR primer was used to introduce silent codon changes (i.e., the amino acid encoded is not changed) at the amino terminus of the protein which did not affect the sequence of the polypeptide. These changes led to increased production of FY2 polymerase. FY3 was designed to promote increased translation efficiency *in vivo*. In addition to the silent codon changes introduced in pREFY2pref, a GCT codon was added in the second position (SEQ. ID. NO. 2), as occurs frequently in strongly expressed genes in *E. coli*. This adds an amino acid to the sequence of FY2, and hence the protein was given its own designation FY3. Both constructs produce more enzyme than pRE273Y.

Silent codon changes such as the following increase protein production in *E. coli*:

substitution of the codon GAG for GAA;

substitution of the codon AGG, AGA, CGG or CGA for CGT or CGC;

substitution of the codon CTT, CTC, CTA, TTG or TTA for CTG; substitution of the codon ATA for ATT or ATC;

substitution of the codon GGG or GGA for GGT or GGC.

The present invention also provides a method for determining the nucleotide base sequence of a DNA molecule. The method includes providing a DNA molecule annealed with a primer molecule able to hybridize to the DNA molecule; and incubating the annealed molecules in a vessel containing at least one deoxynucleotide triphosphate, and a DNA polymerase of the present invention. Also provided is at least one DNA synthesis terminating agent which terminates DNA synthesis at a specific nucleotide base. The method further includes separating the DNA products of the incubating reaction according to size, whereby at least a part of the nucleotide base sequence of the DNA molecule can be determined.

In preferred embodiments, the sequencing is performed at a temperature above 50°C, 60°C, or 70°C.

In other preferred embodiments, the DNA polymerase has less than 1000, 250, 100, 50, 10 or even 2 units of exonuclease activity per mg of polymerase (measured by standard procedure, see below) and is able to utilize primers having only 4, 6 or 10 bases; and the concentration of all four deoxynucleoside triphosphates at the start of the incubating step is sufficient to allow DNA synthesis to continue until terminated by the agent, e.g., a ddNTP.

For cycle sequencing, the DNA polymerases of the present invention make it possible to use significantly lower amounts of dideoxynucleotides compared to naturally occurring enzymes. That is, the method involves providing an excess amount of deoxynucleotides to all four dideoxynucleotides in a cycle sequencing reaction, and performing the cycle sequencing reaction.

5 Preferably, more than 2, 5, 10 or even 100 fold excess of a dNTP is provided to the corresponding ddNTP.

In a related aspect, the invention features a kit or solution for DNA sequencing including a DNA polymerase of the present invention and a reagent necessary for the sequencing such as dITP, deaza GTP, a chain terminating agent such as a ddNTP, and a manganese-containing solution or powder and optionally a pyrophosphatase.

10 In another aspect, the invention features a method for providing a DNA polymerase of the present invention by providing a nucleic acid sequence encoding the modified DNA polymerase, expressing the nucleic acid within a host cell, and purifying the DNA polymerase from the host cell.

15 In another related aspect, the invention features a method for sequencing a strand of DNA essentially as described above with one or more (preferably 2, 3 or 4) deoxyribonucleoside triphosphates, a DNA polymerase of the present invention, and a first chain terminating agent. The DNA polymerase causes the primer to be elongated to form a first series of first DNA products differing in the length of the elongated primer, each first DNA product having a chain terminating agent at its elongated end, and the number of molecules of each first DNA products being approximately the same for substantially all DNA products differing in length by no more than 20 bases. The method also features providing a second chain terminating agent in the hybridized mixture at a concentration different from the first chain terminating agent, wherein the DNA polymerase causes production of a second series of second DNA products differing in the length of the elongated primer, with each second DNA product having the second chain terminating agent at its elongated end. The number of molecules of each second DNA product is approximately the same for substantially all second DNA products differing in length from each other by from 1 to 20 bases, and is distinctly different from the number of molecules of all the first DNA products having a length differing by no more than 20 bases from that of said second DNA products.

20 25 In preferred embodiments, three or four such chain terminating agents can be used to make different products and the sequence reaction is provided with a magnesium ion, or even a manganese or iron ion (e.g., at a concentration between 0.05 and 100 mM, preferably between 1 and 10 mM); and the DNA products are separated according to molecular weight in four or less lanes of a gel.

30 35 In another related aspect, the invention features a method for sequencing a nucleic acid by combining an oligonucleotide primer, a nucleic acid to be sequenced, between one and four deoxyribonucleoside triphosphates, a DNA polymerase of the present invention, and at least two chain terminating agents in different amounts, under conditions favoring extension of the oligonucleotide primer to form nucleic acid fragments complementary to the nucleic acid to be sequenced. For example, the chain terminating agent may be a dideoxynucleotide terminator for adenine, guanine, cytosine or thymine. The method further includes separating the nucleic acid fragments by size and determining the nucleic acid sequence. The agents are differentiated from each other by intensity of a label in the primer extension products.

40 While it is common to use gel electrophoresis to separate DNA products of a DNA sequencing reaction, those in the art will recognize that other methods may also be used. Thus, it is possible to detect each of the different fragments using procedures such as time of flight mass spectrometry, electron microscopy, and single molecule detection methods.

45 The invention also features an automated DNA sequencing apparatus having a reactor including reagents which provide at least two series of DNA products formed from a single primer and a DNA strand. Each DNA product of a series differs in molecular weight and has a chain terminating agent at one end. The reagents include a DNA polymerase of the present invention. The apparatus includes a separating means for separating the DNA product along one axis of the separator to form a series of bands. It also includes a band reading means for determining the position and intensity of each band after separation along the axis, and a computing means that determines the DNA sequence of the DNA strand solely from the position and intensity of the bands along the axis and not from the wavelength of emission of light from any label that may be present in the separating means.

50 Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

#### Description of the Preferred Embodiments

55 The drawings will first briefly be described.

#### Drawings

Figs 1-4 are the DNA sequences, and corresponding amino acid sequences, of FY2, FY3, and the DNA polymer-

ases of T. flavus and Thermus thermophilus, respectively. Figure 5 is the DNA sequence and corresponding amino acid sequence of FY4.

### Examples

The following examples serve to illustrate the DNA polymerases of the present invention and their use in sequencing.

#### Preparation of FY DNA Polymerases (FY2 and FY3)

##### Bacterial Strains

*E. coli* strains: MV1190 [ $\Delta(srl - recA)$  306::Tn10,  $\Delta(lac-proAB)$ , *thi, supE*, F' (*traD36 proAB+ lacI<sup>q</sup> lacZ*  $\Delta M15$ )]; DH $\lambda^+$  [*gyrA96, recA1, relA1, endA1, thi-1, hsdR17, supE44, λ<sup>+</sup>*]; M5248 [ $\lambda(bio275, cl857, clI+$ , N+,  $\Delta(H1)$ )].

##### PCR

Reaction conditions based on the procedure of Barnes (91 Proc. Nat'l. Acad. Sci. 2216-2220, 1994) were as follows: 20mM Tricine pH8.8, 85mM KOAc, 200mM dNTPs, 10% glycerol, 5% DMSO, 0.5mM each primer, 1.5mM MgOAc, 2.5 U HotTub (Amersham Life Science Inc.), 0.025 U DeepVent (New England Biolabs), 1-100 ng target DNA per 100ml reaction. Cycling conditions were 94°C 30s, 68°C 10m40s for 8 cycles; then 94°C 30s, 68°C 12m00s for 8 cycles; then 94°C 30s, 68°C 13m20s for 8 cycles; then 94°C 30s, 68°C 14m40s for 8 cycles.

##### *In vitro* mutagenesis

Restriction enzyme digestions, plasmid preparations, and other *in vitro* manipulations of DNA were performed using standard protocols (Sambrook et al., Molecular Cloning 2nd Ed. Cold Spring Harbor Press, 1989). PCR (see protocol above) was used to introduce a Phe to Tyr amino acid change at codon 667 of native Taq DNA polymerase (which is codon 396 of FY2). Oligonucleotide primer 1 dGCTTGGGCAGAGGATCCGCCGG (SEQ. ID. NO. 3) spans nucleotides 954 to 976 of the coding region of SEQ. ID. NO. 1 including a BamHI restriction site. Mutagenic oligo primer 2 dGGGATGGCTAGCTCCTGGGAGAGGCAGGTGGGCCGACATGCCGTAGA GGACCCCGTAGTTGATGG (SEQ. ID. NO. 4) spans nucleotides 1178 to 1241 including an Nhel site and codon 396 of Sequence ID. NO. 1. A clone of exo-Taq deleted for the first 235 amino acids, pWB253 encoding DeltaTaq polymerase (Barnes, 112 Gene 29-35, 1992) was used as template DNA. Any clone of Taq polymerase or genomic DNA from *Thermus aquaticus* could also be utilized to amplify the identical PCR product. The PCR product was digested with BamHI and Nhel, and this fragment was ligated to BamHI/Nhel digested pWB253 plasmid to replace the corresponding fragment to create pWB253Y, encoding polymerase FY1. Cells of *E. coli* strain MV1190 were used for transformation and induction of protein expression, although any host strain carrying a *lac* repressor could be substituted. DNA sequencing verified the Phe to Tyr change in the coding region.

PCR primer 3 dGGAATTCCATATGGACGATCTGAAGCTCTCC (SEQ. ID. NO. 5) spanning the start codon and containing restriction enzyme sites, was used with PCR primer 4 dGGGGTACCAAGCTTCACTCCTGGCGAGAG (SEQ. ID. NO. 6) containing restriction sites and spanning the stop codon (codon 562 of Sequence ID. NO. 1). A methionine start codon and restriction enzyme recognition sequences were added to PCR primer 5 dGGAATTCCAT-ATGCTGGAGAGGCTTGAGTTT (SEQ. ID. NO. 7), which was used with primer 4 above. PCR was performed using the above primer pairs, and plasmid pWB253Y as template. The PCR products were digested with restriction enzymes NdeI and KpnI and ligated to NdeI/KpnI digested vector pRE2 (Reddi et al., 17 Nucleic Acids Research 10,473-10,488, 1989) to make plasmids pRE236Y, encoding FY1 polymerase, and pRE273Y encoding FY2 polymerase, respectively. Cells of *E. coli* strain DH $\lambda^+$  were used for primary transformation with this and all subsequent pRE2 constructions, and strain M5248 ( $\lambda cl857$ ) was used for protein expression, although any comparable pair of *E. coli* strains carrying the cl<sup>+</sup> and cl857 alleles could be utilized. Alternatively, any *rec<sup>+</sup> cl<sup>+</sup>* strain could be induced by chemical agents such as nalidixic acid to produce the polymerase. The sequences of both genes were verified. pRE273Y was found to produce a single polypeptide band on SDS polyacrylamide gels, unlike pRE253Y or pRE236Y.

Primer 6 dGGAATTCCATATGCTGGAACGTCTGGAGTTGGCAGCCTC CTC (SEQ. ID. NO. 8) and primer 4 were used to make a PCR product introducing silent changes in codon usage of FY2. The product was digested with NdeI/BamHI and ligated to a pRE2 construct containing the 3' end of FY2 to create pREFY2pref, encoding FY2 DNA polymerase. Primer 7 dGGAATTCCATATGGCTCTGGAACGTCTGGAGTTGGCAGCCTC CTC (SEQ. ID. NO. 9) and primer 4 were used to make a PCR product introducing an additional alanine codon commonly occurring at the second position of highly expressed genes. The NdeI/BamHI digested fragment was used as above to create pREFY3, encoding FY3

DNA polymerase.

Preparation of FY4 DNA Polymerase

5 Bacterial Strains

*E. coli* strains: DH1 $\lambda^+$  [*gyrA96, recA1, relA1, endA1, thi-1, hsdR17, supE44, λ<sup>+</sup>*]; M5248 [ $\lambda$  (*bio275, cl857, clI1+, N+, Δ (H1)*)].

10 PCR

Genomic DNA was prepared by standard techniques from *Thermus thermophilus*. The DNA polymerase gene of *Thermus thermophilus* is known to reside on a 3 kilobase AlwNI fragment. To enrich for polymerase sequences in some PCR reactions, the genomic DNA was digested prior to PCR with AlwNI, and fragments of approximately 3 kb were selected by agarose gel electrophoresis to be used as template DNA. Reaction conditions were as follows: 10mM Tris pH8.3, 50mM KCl, 800μM dNTPs, 0.001% gelatin, 1.0μM each primer, 1.5mM MgCl<sub>2</sub>, 2.5 U Tth, 0.025 U Deepvent (New England Biolabs), per 100μl reaction. Cycling conditions were 94°C 2 min, then 35 cycles of 94°C 30s, 55°C 30s, 72°C 3 min, followed by 72 °C for 7 min.

20 *In vitro* mutagenesis

Restriction enzyme digestions, plasmid preparations, and other *in vitro* manipulations of DNA were performed using standard protocols (Sambrook et al., 1989). Plasmid pMR1 was constructed to encode an exonuclease-free polymerase, with optimized codons for expression in *E. coli* at the 5' end. Primer 8 (SEQ. ID. NO. 10) (GGAATTCAT-ATGCTGGAACGTCTGGATTCTGGCAGCCTC) was used with Primer 9 (SEQ. ID. NO. 11) (GGGGTACCCCTAACCCCTT-GGCGGAAAGGCCAGTC) to create a PCR product from *Tth* genomic DNA, which was digested with restriction enzymes NdeI and KpnI and inserted into plasmid pRE2 (Reddi et al., 1989, *Nucleic Acids Research* 17, 10473 - 10488) digested with the same enzymes.

To create the desired F396Y mutation, two PCR products were made from *Tth* chromosomal DNA. Primer 8 above was used in combination with Primer 10 (SEQ. ID. NO. 12) (GGGATGGCTAGCTCCTGGGAGAGCCTAT-GGGCGGACAT GCCGTAGAGGACGCCGTAGTTCACCG) to create a portion of the gene containing the F to Y amino acid change as well as a silent change to create an NheI restriction site. Primer 11 (SEQ. ID. NO. 13) (CTAGCTAGCCATCCCCTA CGAAGAACGGTGGCCT) was used in combination with primer 9 above to create a portion of the gene from the introduced NheI site to the stop codon at the 3' end of the coding sequence. The PCR product of Primers 8 and 10 was digested with NdeI and NheI, and the PCR product of Primers 9 and 11 was digested with NheI and KpnI. These were introduced into expression vector pRE2 which was digested with NdeI and KpnI to produce plasmid pMR5. In addition to the desired changes, pMR5 was found to have a spurious change introduced by PCR, which led to an amino acid substitution, K234R. Plasmid pMR8 was created to eliminate this substitution, by replacing the AfIII/BamHI fragment of pMR5 for the corresponding fragment from pMR1. The FY4 polymerase encoded by plasmid pMR8 (SEQ. ID. NO. 14) is given in Figure 5.

Cells of *E. coli* strain DH1 $\lambda^+$  were used for primary transformation, and strain M5248 ( $\lambda$ cl857) was used for protein expression, although any comparable pair of *E. coli* strains carrying the cl<sup>+</sup> and cl857 alleles could be utilized. Alternatively, any *rec*<sup>+</sup> cl<sup>+</sup> strain could be induced by chemical agents such as nalidixic acid to produce the polymerase.

45 Protein Sequencing

Determinations of amino terminal protein sequences were performed at the W.M. Keck Foundation, Biotechnology Resource Laboratory, New Haven, Connecticut.

50 Purification of Polymerases

A 1 liter culture of 2X LB (2% Bacto-Tryptone, 1% Bacto-Yeast Extract, 0.5% NaCl) + 0.2% Casamino Acids + 20 mM KPO<sub>4</sub> pH 7.5 + 50 μg/ml Ampicillin was inoculated with a glycerol stock of the appropriate cell strain and grown at 30°C with agitation until cells were in log phase (0.7-1.0 OD<sub>590</sub>). 9 liters of 2X LB + 0.2% Casamino Acids + 20 mM KPO<sub>4</sub> pH 7.5 + 0.05% Mazu Anti-loam was inoculated with 1 liter of log phase cells in 10 liter Microferm Fermentors (New Brunswick Scientific Co.). Cells were grown at 30°C under 15.psi pressure, 350-450 rpm agitation, and an air flow rate of 14,000 cc/min ±1000 cc/min. When the OD<sub>590</sub> reached 1.5-2.0, the cultures were induced by increasing the temperature to 40-42°C for 90-120 minutes. The cultures were then cooled to < 20°C and the cells harvested by

centrifugation in a Sorvall RC-3B centrifuge at 5000 rpm at 4°C for 15-20 minutes. Harvested cells were stored at -80°C.

Frozen cells were broken into small pieces and resuspended in pre-warmed (90-95°C) Lysis Buffer (20 mM Tris pH 8.5, 1 mM EDTA, 10 mM MgCl<sub>2</sub>, 16 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.1% Tween 20, 0.1% Nonidet P-40, 1 mM PMSF). Resuspended cells were then heated rapidly to 80°C and incubated at 80°C for 20 minutes with constant stirring. The suspension was then rapidly cooled on ice. The cell debris was removed by centrifugation using a Sorvall GSA rotor at 10,000 rpm for 20 minutes at 4°C. The NaCl concentration of the supernatant was adjusted to 300 mM. The sample was then passed through a diethylaminoethyl cellulose (Whatman DE-52) column that had been previously equilibrated with Buffer A (20 mM Tris pH 8.5, 1 mM EDTA, 0.1% Tween 20, 0.1% Nonidet P-40, 300 mM NaCl, 10% glycerol, 1 mM DTT), and polymerase collected in the flow through. The sample was then diluted to a concentration of NaCl of 100mM and applied to a Heparin-sepharose column. The polymerase was eluted from the column with a NaCl gradient (100-500 mM NaCl). The sample was then dialyzed against Buffer B (20 mM Tris pH 8.5, 1 mM EDTA, 0.1% Tween 20, 0.1% Nonidet P-40, 10 mM KCl, 10% glycerol, 1 mM DTT) and further diluted as needed to lower the conductivity of the sample to the conductivity of Buffer B. The sample was then applied to a diethylaminoethyl (Waters DEAE 15 HR) column and eluted with a 10-500 mM KCl gradient. The polymerase was then diluted with an equal volume of Final Buffer (20 mM Tris pH 8.5, 0.1 mM EDTA, 0.5% Tween 20, 0.5% Nonidet P-40, 100 mM KCl, 50% glycerol, 1 mM DTT) and dialyzed against Final Buffer.

#### Assay of Exonuclease Activity

The exonuclease assay was performed by incubating 5  $\mu$ l (25-150 units) of DNA polymerase with 5  $\mu$ g of labelled [<sup>3</sup>H]-pBR322 PCR fragment ( $1.6 \times 10^4$  cpm/ $\mu$ g DNA) in 100  $\mu$ l of reaction buffer of 20 mM Tris-HCl pH 8.5, 5 mM MgCl<sub>2</sub>, 10 mM KCl, for 1 hour at 60 °C. After this time interval, 200  $\mu$ l of 1:1 ratio of 50  $\mu$ g/ml salmon sperm DNA with 2 mM EDTA and 20% TCA with 2% sodium pyrophosphate were added into the assay aliquots. The aliquots were put on ice for 10 min and then centrifuged at 12,000g for 10 min. Acid-soluble radioactivity in 200  $\mu$ l of the supernatant was quantitated by liquid scintillation counting. One unit of exonuclease activity was defined as the amount of enzyme that catalyzed the acid solubilization of 10 nmol of total nucleotide in 30 min at 60 °C.

#### Utility in DNA Sequencing

##### Example 1: DNA Sequencing with FY Polymerases (e.g., FY2 and FY3)

The following components were added to a microcentrifuge vial (0.5 ml): 0.4 pmol M13 DNA (e.g., M13mp18, 1.0  $\mu$ g); 2  $\mu$ l Reaction Buffer (260 mM Tris-HCl, pH 9.5 65 mM MgCl<sub>2</sub>); 2  $\mu$ l of labeling nucleotide mixture (1.5  $\mu$ M each of dGTP, dCTP and dTTP); 0.5  $\mu$ l (5  $\mu$ Ci) of [ $\alpha$ -<sup>33</sup>P]dATP (about 2000Ci/mmol); 1  $\mu$ l -40 primer (0.5  $\mu$ M; 0.5 pmol/ $\mu$ l 5'GTTTCCCAGTCACGAC-3'); 2  $\mu$ l of a mixture containing 4 U/ $\mu$ l FY polymerase and 6.6 U/ml *Thermoplasma acidophilum* inorganic pyrophosphatase (32 U/ $\mu$ l polymerase and 53 U/ml pyrophosphatase in 20 mM Tris (pH8.5), 100 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5% NP-40, 0.5% TWEEN-20 and 50% glycerol, diluted 8 fold in dilution buffer (10 mM Tris-HCl pH8.0, 1 mM 2-mercaptoethanol, 0.5% TWEEN-20, 0.5% NP-40)); and water to a total volume of 17.5  $\mu$ l. These components (the labeling reaction) were mixed and the vial was placed in a constant-temperature water bath at 45°C for 5 minutes.

Four vials were labeled A, C, G, and T, and filled with 4  $\mu$ l of the corresponding termination mix: ddA termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddATP); ddT termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddTTP); ddC termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddCTP); ddG termination mix (150  $\mu$ M each dATP, dCTP, dGTP, dTTP, 1.5  $\mu$ M ddGTP).

The labeling reaction was divided equally among the four termination vials (4  $\mu$ l to each termination reaction vial), and tightly capped.

The four vials were placed in a constant-temperature water bath at 72°C for 5 minutes. Then 4  $\mu$ l of Stop Solution (95% Formamide 20 mM EDTA, 0.05% Bromophenol Blue, 0.05% Xylene Cyanol FF) added to each vial, and heated briefly to 70°-80°C immediately prior to loading on a sequencing gel (8% acrylamide, 8.3 M urea). Autoradiograms required an 18-36 hour exposure using Kodak XAR-5 film or Amersham Hyperfilm MP. High-quality sequence results with uniform band intensities were obtained. The band intensities were much more uniform than those obtained with similar protocols using Taq DNA polymerase or  $\Delta$ Taq DNA polymerase.

##### Example 2: DNA Cycle Sequencing with FY Polymerases

The following components were added to a microcentrifuge vial (0.5 ml) which is suitable for insertion into a thermocycler machine (e.g., Perkin-Elmer DNA Thermal Cycler): 0.05 pmol or more M13 DNA (e.g., M13mp18, 0.1  $\mu$ g), or 0.1  $\mu$ g double-stranded plasmid DNA (e.g., pUC19); 2  $\mu$ l Reaction Buffer (260 mM Tris-HCl, pH 9.5 65 mM

MgCl<sub>2</sub>); 1 µl 3.0 µM dGTP; 1 µl 3.0 µM dTTP; 0.5 µl (5 µCi) of [ $\alpha$ -<sup>33</sup>P]dATP (about 2000Ci/mmol); 1 µl -40 primer (0.5 µM; 0.5 pmol/µl 5'-GTTTCCCAGTCACGAC-3'); 2 µl of a mixture containing 4 U/µl FY polymerase and 6.6 U/ml *Thermoplasma acidophilum* inorganic pyrophosphatase (32 U/µl polymerase and 53 U/ml pyrophosphatase in 20 mM Tris (pH8.5), 100 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5% NP-40, 0.5% TWEEN-20 and 50% glycerol, diluted 8 fold in dilution buffer (10 mM Tris-HCl pH8.0, 1 mM 2-mercaptoethanol, 0.5% TWEEN-20, 0.5% NP-40)); and water to a total volume of 17.5 µl.

These components (labeling reaction mixture) were mixed and overlaid with 10 µl light mineral oil (Amersham). The vial was placed in the thermocycler and 30-100 cycles (more than 60 cycles is unnecessary) from 45°C for 1 minute to 95°C for 0.5 minute performed. (Temperatures can be cycled from 55°-95°C, if desired). The temperatures may be adjusted if the melting temperature of the primer/template is significantly higher or lower, but these temperatures work well for most primer/templates combinations. This step can be completed in about 3 minutes per cycle.

Four vials were labeled A, C, G, and T, and filled with 4 ml of the corresponding termination mix: ddA termination mix (150 µM each dATP, dCTP, dGTP, dTTP, 1.5 µM ddATP); ddT termination mix (150 µM each dATP, dCTP, dGTP, dTTP, 1.5 µM ddTTP); ddC termination mix (150 µM each dATP, dCTP, dGTP, dTTP, 1.5 µM ddCTP); ddG termination mix (150 µM each dATP, dCTP, dGTP, dTTP, 1.5 µM ddGTP). No additional enzyme is added to the termination vials. The enzyme carried in from the prior (labeling) step is sufficient.

The cycled labeling reaction mixture was divided equally among the four termination vials (4 µl to each termination reaction vial), and overlaid with 10 µl of light mineral oil.

The four vials were placed in the thermocycler and 30-200 cycles (more than 60 cycles is unnecessary) performed from 95°C for 15 seconds, 55°C for 30 seconds, and 72°C for 120 seconds. This step was conveniently completed overnight. Other times and temperatures are also effective.

Six µl of reaction mixture was removed (avoiding oil), 3 µl of Stop Solution (95% Formamide 20 mM EDTA, 0.05% Bromophenol Blue, 0.05% Xylene Cyanol FF) added, and heated briefly to 70°-80°C immediately prior to loading on a sequencing gel. Autoradiograms required an 18-36 hour exposure using Kodak XAR-5 film or Amersham Hyperfilm MP. High-quality sequence results with uniform band intensities were obtained. The band intensities were much more uniform than those obtained with similar protocols using Taq DNA polymerase or  $\Delta$ Taq DNA polymerase.

#### Example 3: Sequencing with dGTP analogs to eliminate compression artifacts.

For either of the sequencing methods outlined in examples 1 and 2, 7-Deaza-2'-deoxy-GTP can be substituted for dGTP in the labeling and termination mixtures at exactly the same concentration as dGTP. When this substitution is made, secondary structures on the gels are greatly reduced. Similarly, 2'-deoxyinosinetriphosphate can also be substituted for dGTP but its concentration must be 10-fold higher than the corresponding concentration of dGTP. Substitution of dITP for dGTP is even more effective in eliminating compression artifacts than 7-deaza-dGTP.

#### Example 4: Other Sequencing methods using FY polymerases

FY polymerases have been adapted for use with many other sequencing methods, including the use of fluorescent primers and fluorescent-dideoxy-terminators for sequencing with the ABI 373A DNA sequencing instrument.

#### Example 5: SDS-Polyacrylamide Gel Electrophoresis

Protein samples were run on a 14 X 16 mm 7.5 or 10% polyacrylamide gel. (Gels were predominantly 10% Polyacrylamide using a 14 X 16 mm Hoefer apparatus. Other sizes, apparatuses, and percentage gels are acceptable. Similar results can also be obtained using the Pharmacia Phast Gel system with SDS, 8-25% gradient gels. Reagent grade and ultrapure grade reagents were used.) The stacking gel consisted of 4% acrylamide (30:0.8, acrylamide: bisacrylamide), 125 mM Tris-HCl pH 6.8, 0.1% Sodium Dodecyl Sulfate (SDS). The resolving gel consisted of 7.5 or 10% acrylamide (30:0.8, acrylamide: bisacrylamide), 375 mM Tris-HCl pH 8.8, 0.1% SDS. Running Buffer consisted of 25 mM Tris, 192 mM Glycine and 0.1% SDS. 1X Sample Buffer consisted of 25 mM Tris-HCl pH 6.8, 0.25% SDS, 10% Glycerol, 0.1M Dithiothreitol, 0.1% Bromophenol Blue, and 1mM EDTA. A 1/4 volume of 5X Sample Buffer was added to each sample. Samples were heated in sample buffer to 90-100°C for approximately 5 minutes prior to loading. A 1.5 mm thick gel was run at 50-100 mA constant current for 1-3 hours (until bromophenol blue was close to the bottom of the gel). The gel was stained with 0.025% Coomassie Blue R250 in 50% methanol, 10% acetic acid and destained in 5% methanol, 7% acetic acid solution. A record of the gel was made by taking a photograph of the gel, by drying the gel between cellulose film sheets, or by drying the gel onto filter paper under a vacuum.

Other embodiments are within the following claims.

## SEQUENCE LISTING

## 5 (1) GENERAL INFORMATION:

(i) APPLICANT: AMERSHAM LIFE SCIENCE

10 (ii) TITLE OF INVENTION: THERMOSTABLE DNA  
POLYMERASES

15 (iii) NUMBER OF SEQUENCES: 14

## 20 (iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Lyon & Lyon  
25 (B) STREET: 633 West Fifth Street  
Suite 4700  
(C) CITY: Los Angeles  
(D) STATE: California  
(E) COUNTRY: U.S.A.  
30 (F) ZIP: 90071-2066

## 35 (v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 3.5" Diskette, 1.44 Mb  
storage  
40 (B) COMPUTER: IBM Compatible  
(C) OPERATING SYSTEM: IBM P.C. DOS 5.0  
(D) SOFTWARE: Word Perfect 5.1

## 45 (vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: To Be Assigned  
(B) FILING DATE:  
50 (C) CLASSIFICATION:

## (vii) PRIOR APPLICATION DATA:

Prior applications total,  
including application  
described below: one

5

- (A) APPLICATION NUMBER: US 08/455,686  
(B) FILING DATE: May 31, 1995

10

(viii) ATTORNEY/AGENT INFORMATION:

15

- (A) NAME: Warburg, Richard J.  
(B) REGISTRATION NUMBER: 32,327  
(C) REFERENCE/DOCKET NUMBER: 219/304-PCT

20

(ix) TELECOMMUNICATION INFORMATION:

25

- (A) TELEPHONE: (213) 489-1600  
(B) TELEFAX: (213) 955-0440  
(C) TELEX: 67-3510

25

(2) INFORMATION FOR SEQ ID NO: 1:

30

(i) SEQUENCE CHARACTERISTICS:

35

- (A) LENGTH: 1686 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

40

(ix) FEATURE:

45

- (A) NAME/KEY: FY2  
(B) LOCATION: 1...1683

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATG CTG GAG AGG CTT GAG TTT GGC AGC CTC CTC CAC GAG TTC GGC CTT  
Met Leu Glu Arg Leu Glu Phe Gly Ser Leu Leu His Glu Phe Gly Leu  
1 5 10 15

48

CTG GAA AGC CCC AAG GCC CTG GAG GAG GCC CCC TGG CCC CCG CCG GAA  
Leu Glu Ser Pro Lys Ala Leu Glu Ala Pro Trp Pro Pro Pro Glu  
20 25 30

96

55

GGG GCC TTC GTG GGC TTT GTG CTT TCC CGC AAG GAG CCC ATG TGG GCC 144

	Gly Ala Phe Val Gly Phe Val Leu Ser Arg Lys Glu Pro Met Trp Ala		
	35	40	45
5	GAT CTT CTG GCC CTG GCC GCG GCC AGG GGG GGC CGG GTC CAC CGG GCC		192
	Asp Leu Leu Ala Leu Ala Ala Arg Gly Gly Arg Val His Arg Ala		
	50	55	60
10	CCC GAG CCT TAT AAA GCC CTC AGG GAC CTG AAG GAG GCG CGG GGG CTT		240
	Pro Glu Pro Tyr Lys Ala Leu Arg Asp Leu Lys Glu Ala Arg Gly Leu		
	65	70	75
	CTC GCC AAA GAC CTG AGC GTT CTG GCC CTG AGG GAA GGC CTT GGC CTC		288
15	Leu Ala Lys Asp Leu Ser Val Leu Ala Leu Arg Glu Gly Leu Gly Leu		
	85	90	95
20	CCG CCC GGC GAC GAC CCC ATG CTC CTC GCC TAC CTC CTG GAC CCT TCC		336
	Pro Pro Gly Asp Asp Pro Met Leu Leu Ala Tyr Leu Leu Asp Pro Ser		
	100	105	110
	AAC ACC ACC CCC GAG GGG GTG GCC CGG CGC TAC GGC GGG GAG TGG ACG		384
	Asn Thr Thr Pro Glu Gly Val Ala Arg Arg Tyr Gly Gly Glu Trp Thr		
	115	120	125
25	GAG GAG GCG GGG GAG CGG GCC CTT TCC GAG AGG CTC TTC GCC AAC		432
	Glu Glu Ala Gly Glu Arg Ala Ala Leu Ser Glu Arg Leu Phe Ala Asn		
	130	135	140
30	CTG TGG GGG AGG CTT GAG GGG GAG GAG AGG CTC CTT TGG CTT TAC CGG		480
	Leu Trp Gly Arg Leu Glu Gly Glu Arg Leu Leu Trp Leu Tyr Arg		
	145	150	155
35	GAG GTG GAG AGG CCC CTT TCC GCT GTC CTG GCC CAC ATG GAG GCC ACG		528
	Glu Val Glu Arg Pro Leu Ser Ala Val Leu Ala His Met Glu Ala Thr		
	165	170	175
40	GGG GTG CGC CTG GAC GTG GCC TAT CTC AGG GCC TTG TCC CTG GAG GTG		576
	Gly Val Arg Leu Asp Val Ala Tyr Leu Arg Ala Leu Ser Leu Glu Val		
	180	185	190
45	GCC GAG GAG ATC GCC CGC CTC GAG GCC GAG GTC TTC CGC CTG GCC GGC		624
	Ala Glu Glu Ile Ala Arg Leu Glu Ala Glu Val Phe Arg Leu Ala Gly		
	195	200	205
50	CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AGG GTC CTC TTT		672
	His Pro Phe Asn Leu Asn Ser Arg Asp Gln Leu Glu Arg Val Leu Phe		
	210	215	220
55	GAC GAG CTA GGG CTT CCC GCC ATC GGC AAG ACG GAG AAG ACC GGC AAG		720
	Asp Glu Leu Gly Leu Pro Ala Ile Gly Lys Thr Glu Lys Thr Gly Lys		
	225	230	235
	240		

## EP 0 745 676 A1

CGC TCC ACC AGC GCC GCC GTC CTG GAG GCC CTC CGC GAG GCC CAC CCC Arg Ser Thr Ser Ala Ala Val Leu Glu Ala Leu Arg Glu Ala His Pro 245	250	255	768
5 ATC GTG GAG AAG ATC CTG CAG TAC CGG GAG CTC ACC AAG CTG AAG AGC Ile Val Glu Lys Ile Leu Gln Tyr Arg Glu Leu Thr Lys Leu Lys Ser 260	265	270	816
10 ACC TAC ATT GAC CCC TTG CCG GAC CTC ATC CAC CCC AGG ACG GGC CGC Thr Tyr Ile Asp Pro Leu Pro Asp Leu Ile His Pro Arg Thr Gly Arg 275	280	285	864
15 CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GGC AGG CTA AGT Leu His Thr Arg Phe Asn Gln Thr Ala Thr Ala Thr Gly Arg Leu Ser 290	295	300	912
20 AGC TCC GAT CCC AAC CTC CAG AAC ATC CCC GTC CGC ACC CCG CTT GGG Ser Ser Asp Pro Asn Leu Gln Asn Ile Pro Val Arg Thr Pro Leu Gly 305	310	315	960
25 CAG AGG ATC CGC CGG GCC TTC ATC GCC GAG GAG GGG TGG CTA TTG GTG Gln Arg Ile Arg Arg Ala Phe Ile Ala Glu Glu Gly Trp Leu Leu Val 325	330	335	1008
30 GCC CTG GAC TAT AGC CAG ATA GAG CTC AGG GTG CTG GCC CAC CTC TCC Ala Leu Asp Tyr Ser Gln Ile Glu Leu Arg Val Leu Ala His Leu Ser 340	345	350	1056
35 GGC GAC GAG AAC CTG ATC CGG GTC TTC CAG GAG GGG CGG GAC ATC CAC Gly Asp Glu Asn Leu Ile Arg Val Phe Gln Glu Gly Arg Asp Ile His 355	360	365	1104
40 ACG GAG ACC GCC AGC TGG ATG TTC GGC GTC CCC CGG GAG GCC GTG GAC Thr Glu Thr Ala Ser Trp Met Phe Gly Val Pro Arg Glu Ala Val Asp 370	375	380	1152
45 CCC CTG ATG CGC CGG GCG GCC AAG ACC ATC AAC TAC GGG GTC CTC TAC Pro Leu Met Arg Arg Ala Ala Lys Thr Ile Asn Tyr Gly Val Leu Tyr 385	390	395	1200
50 GGC ATG TCG GCC CAC CGC CTC TCC CAG GAG CTA GCC ATC CCT TAC GAG Gly Met Ser Ala His Arg Leu Ser Gln Glu Leu Ala Ile Pro Tyr Glu 405	410	415	1248
GAG GCC CAG GCC TTC ATT GAG CGC TAC TTT CAG AGC TTC CCC AAG GTG Glu Ala Gln Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys Val 420	425	430	1296
55 CGG GCC TGG ATT GAG AAG ACC CTG GAG GAG GGC AGG AGG CGG GGG TAC Arg Ala Trp Ile Glu Lys Thr Leu Glu Glu Gly Arg Arg Arg Gly Tyr 435	440	445	1344

GTG GAG ACC CTC TTC GGC CGC CGC TAC GTG CCA GAC CTA GAG GCC 1392  
 Val Glu Thr Leu Phe Gly Arg Arg Arg Tyr Val Pro Asp Leu Glu Ala  
 450 455 460

CGG GTG AAG AGC GTG CGG GAG GCG GCC GAG CGC ATG GCC TTC AAC ATG 1440  
 Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn Met  
 465 470 475 480

CCC GTC CAG GGC ACC GCC GAC CTC ATG AAG CTG GCT ATG GTG AAG 1488  
 Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val Lys  
 485 490 495

CTC TTC CCC AGG CTG GAG GAA ATG GGG GCC AGG ATG CTC CTT CAG GTC 1536  
 Leu Phe Pro Arg Leu Glu Glu Met Gly Ala Arg Met Leu Leu Gln Val  
 500 505 510

CAC GAC GAG CTG GTC CTC GAG GCC CCA AAA GAG AGG GCG GAG GCC GTG 1584  
 His Asp Glu Leu Val Leu Glu Ala Pro Lys Glu Arg Ala Glu Ala Val  
 515 520 525

GCC CGG CTG GCC AAG GAG GTC ATG GAG GGG GTG TAT CCC CTG GCC GTG 1632  
 Ala Arg Leu Ala Lys Glu Val Met Glu Gly Val Tyr Pro Leu Ala Val  
 530 535 540

CCC CTG GAG GTG GAG GTG GGG ATA GGG GAG GAC TGG CTC TCC GCC AAG 1680  
 Pro Leu Glu Val Glu Val Gly Ile Gly Glu Asp Trp Leu Ser Ala Lys  
 545 550 555 560

GAG TGA 1686  
 Glu \*

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1689 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: FY3
- (B) LOCATION: 1...1686

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

ATG GCT CTG GAA CGT CTG GAG TTT GGC AGC CTC CTC CAC GAG TTC GGC 48

	Met Ala Leu Glu Arg Leu Glu Phe Gly Ser Leu Leu His Glu Phe Gly			
1	5	10	15	
5	CTT CTG GAA AGC CCC AAG GCC CTG GAG GAG GCC CCC TGG CCC CCG CCG Leu Leu Glu Ser Pro Lys Ala Leu Glu Ala Pro Trp Pro Pro Pro		96	
	20	25	30	
10	GAA GGG GCC TTC GTG GGC TTT GTG CTT TCC CGC AAG GAG CCC ATG TGG Glu Gly Ala Phe Val Gly Phe Val Leu Ser Arg Lys Glu Pro Met Trp		144	
	35	40	45	
15	GCC GAT CTT CTG GCC CTG GCC GCC AGG GGG GGC CGG GTC CAC CGG Ala Asp Leu Leu Ala Leu Ala Ala Arg Gly Gly Arg Val His Arg		192	
	50	55	60	
20	GCC CCC GAG CCT TAT AAA GCC CTC AGG GAC CTG AAG GAG GCG CGG GGG Ala Pro Glu Pro Tyr Lys Ala Leu Arg Asp Leu Lys Glu Ala Arg Gly		240	
	65	70	75	80
25	CTT CTC GCC AAA GAC CTG AGC GTT CTG GCC CTG AGG GAA GGC CTT GGC Leu Leu Ala Lys Asp Leu Ser Val Leu Ala Leu Arg Glu Gly Leu Gly		288	
	85	90	95	
30	CTC CCG CCC GGC GAC CCC ATG CTC CTC GCC TAC CTC CTG GAC CCT Leu Pro Pro Gly Asp Asp Pro Met Leu Leu Ala Tyr Leu Leu Asp Pro		336	
	100	105	110	
35	TCC AAC ACC ACC CCC GAG GGG GTG GCC CGG CGC TAC GGC GGG GAG TGG Ser Asn Thr Thr Pro Glu Gly Val Ala Arg Arg Tyr Gly Gly Glu Trp		384	
	115	120	125	
40	ACG GAG GAG GCG GGG GAG CGG GCC CTT TCC GAG AGG CTC TTC GCC Thr Glu Ala Gly Glu Arg Ala Ala Leu Ser Glu Arg Leu Phe Ala		432	
	130	135	140	
45	AAC CTG TGG GGG AGG CTT GAG GGG GAG GAG AGG CTC CTT TGG CTT TAC Asn Leu Trp Gly Arg Leu Glu Gly Glu Arg Leu Leu Trp Leu Tyr		480	
	145	150	155	160
50	CGG GAG GTG GAG AGG CCC CTT TCC GCT GTC CTG GCC CAC ATG GAG GCC Arg Glu Val Glu Arg Pro Leu Ser Ala Val Leu Ala His Met Glu Ala		528	
	165	170	175	
55	ACG GGG GTG CGC CTG GAC GTG GCC TAT CTC AGG GCC TTG TCC CTG GAG Thr Gly Val Arg Leu Asp Val Ala Tyr Leu Arg Ala Leu Ser Leu Glu		576	
	180	185	190	
60	GTG GCC GAG GAG ATC GCC CGC CTC GAG GCC GAG GTC TTC CGC CTG GCC Val Ala Glu Glu Ile Ala Arg Leu Glu Ala Glu Val Phe Arg Leu Ala		624	
	195	200	205	
65	GGC CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AGG GTC CTC		672	

	Gly His Pro Phe Asn Leu Asn Ser Arg Asp Gln Leu Glu Arg Val Leu		
210	215	220	
5	TTT GAC GAG CTA GGG CTT CCC GCC ATC GGC AAG ACG GAG AAG ACC GGC	720	
	Phe Asp Glu Leu Gly Leu Pro Ala Ile Gly Lys Thr Glu Lys Thr Gly		
225	230	235	
10	AAG CGC TCC ACC AGC GCC GCC GTC CTG GAG GCC CTC CGC GAG GCC CAC	768	
	Lys Arg Ser Thr Ser Ala Ala Val Leu Glu Ala Leu Arg Glu Ala His		
	245	250	255
15	CCC ATC GTG GAG AAG ATC CTG CAG TAC CGG GAG CTC ACC AAG CTG AAG	816	
	Pro Ile Val Glu Lys Ile Leu Gln Tyr Arg Glu Leu Thr Lys Leu Lys		
	260	265	270
20	AGC ACC TAC ATT GAC CCC TTG CCG GAC CTC ATC CAC CCC AGG ACG GGC	864	
	Ser Thr Tyr Ile Asp Pro Leu Pro Asp Leu Ile His Pro Arg Thr Gly		
	275	280	285
25	CGC CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GCC ACG GGC AGG CTA	912	
	Arg Leu His Thr Arg Phe Asn Gln Thr Ala Thr Gly Arg Leu		
	290	295	300
30	AGT AGC TCC GAT CCC AAC CTC CAG AAC ATC CCC GTC CGC ACC CCG CTT	960	
	Ser Ser Ser Asp Pro Asn Leu Gln Asn Ile Pro Val Arg Thr Pro Leu		
	305	310	315
35	GGG CAG AGG ATC CGC CGG GCC TTC ATC GCC GAG GAG GGG TGG CTA TTG	1008	
	Gly Gln Arg Ile Arg Arg Ala Phe Ile Ala Glu Glu Gly Trp Leu Leu		
	325	330	335
40	GTG GCC CTG GAC TAT AGC CAG ATA GAG CTC AGG GTG CTG GCC CAC CTC	1056	
	Val Ala Leu Asp Tyr Ser Gln Ile Glu Leu Arg Val Leu Ala His Leu		
	340	345	350
45	TCC GGC GAC GAG AAC CTG ATC CGG GTC TTC CAG GAG GGG CGG GAC ATC	1104	
	Ser Gly Asp Glu Asn Leu Ile Arg Val Phe Gln Glu Gly Arg Asp Ile		
	355	360	365
50	CAC ACG GAG ACC GCC AGC TGG ATG TTC GGC GTC CCC CGG GAG GCC GTG	1152	
	His Thr Glu Thr Ala Ser Trp Met Phe Gly Val Pro Arg Glu Ala Val		
	370	375	380
55	GAC CCC CTG ATG CGC CGG GCG AAG ACC ATC AAC TAC GGG GTC CTC	1200	
	Asp Pro Leu Met Arg Arg Ala Ala Lys Thr Ile Asn Tyr Gly Val Leu		
	385	390	395
	TAC GGC ATG TCG GCC CAC CGC CTC TCC CAG GAG CTA GCC ATC CCT TAC	1248	
	Tyr Gly Met Ser Ala His Arg Leu Ser Gln Glu Leu Ala Ile Pro Tyr		
	405	410	415

GAG GAG GCC CAG GCC TTC ATT GAG CGC TAC TTT CAG AGC TTC CCC AAG 1296  
 Glu Glu Ala Gln Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys  
 420 425 430

GTG CGG GCC TGG ATT GAG AAG ACC CTG GAG GAG GGC AGG AGG CGG GGG 1344  
 Val Arg Ala Trp Ile Glu Lys Thr Leu Glu Gly Arg Arg Arg Gly  
 435 440 445

TAC GTG GAG ACC CTC TTC GGC CGC CGC TAC GTG CCA GAC CTA GAG 1392  
 Tyr Val Glu Thr Leu Phe Gly Arg Arg Arg Tyr Val Pro Asp Leu Glu  
 450 455 460

GCC CGG GTG AAG AGC GTG CGG GAG GCG GCC GAG CGC ATG GCC TTC AAC 1440  
 Ala Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn  
 465 470 475 480

ATG CCC GTC CAG GGC ACC GCC GAC CTC ATG AAG CTG GCT ATG GTG 1488  
 Met Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val  
 485 490 495

AAG CTC TTC CCC AGG CTG GAG GAA ATG GGG GCC AGG ATG CTC CTT CAG 1536  
 Lys Leu Phe Pro Arg Leu Glu Glu Met Gly Ala Arg Met Leu Leu Gln  
 500 505 510

GTC CAC GAC GAG CTG GTC CTC GAG GCC CCA AAA GAG AGG GCG GAG GCC 1584  
 Val His Asp Glu Leu Val Leu Glu Ala Pro Lys Glu Arg Ala Glu Ala  
 515 520 525

GTG GCC CGG CTG GCC AAG GAG GTC ATG GAG GGG GTG TAT CCC CTG GCC 1632  
 Val Ala Arg Leu Ala Lys Glu Val Met Glu Gly Val Tyr Pro Leu Ala  
 530 535 540

GTG CCC CTG GAG GTG GAG GTG GGG ATA GGG GAG GAC TGG CTC TCC GCC 1680  
 Val Pro Leu Glu Val Glu Val Gly Ile Gly Glu Asp Trp Leu Ser Ala  
 545 550 555 560

AAG GAG TGA 1689  
 Lys Glu \*

## (2) INFORMATION FOR SEQ ID NO: 3:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GCTTGGGCAG AGGATCCGCC GGG

23

5

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 64 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

20 GGGATGGCTA GCTCCTGGGA GAGGCGGTGG GCCGACATGC CGTAGAGGAC  
CCCGTAGTTG ATGG

50

64

25 (2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GGAATTCCAT ATGGACGATC TGAAGCTCTC C

31

40

(2) INFORMATION FOR SEQ ID NO: 6:

45

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

55

GGGGTACCAA GCTTCACTCC TTGGCGGAGA G

31

5

(2) INFORMATION FOR SEQ ID NO: 7:

10

(i) SEQUENCE CHARACTERISTICS:

15

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

20

GGAATTCCAT ATGCTGGAGA GGCTTGAGTT T

31

25

(2) INFORMATION FOR SEQ ID NO: 8:

30

(i) SEQUENCE CHARACTERISTICS:

35

- (A) LENGTH: 43 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

40

GGAATTCCAT ATGCTGGAAC GTCTGGAGTT TGGCAGCCTC CTC

43

45

(2) INFORMATION FOR SEQ ID NO: 9:

50

(i) SEQUENCE CHARACTERISTICS:

55

- (A) LENGTH: 46 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

55

GGAATTCCAT ATGGCTCTGG AACGTCTGGA GTTTGGCAGC CTCCTC

46

5 (2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- 10 (A) LENGTH: 40 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

20 GGAATTCCAT ATGCTGGAAC GTCTGGAATT CGGCAGCCTC

40

25 (2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 32 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

40 GGGGTACCCCT AACCCCTTGGC GGAAAGCCAG TC

32

45 (2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- 50 (A) LENGTH: 64 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

55 GGGATGGCTA GCTCCTGGGA GAGCCTATGG GCGGACATGC CGTAGAGGAC

50

GCCGTAGTTC ACCG

64

## 5 (2) INFORMATION FOR SEQ ID NO: 13:

## (i) SEQUENCE CHARACTERISTICS:

- 10 (A) LENGTH: 35 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

20 CTAGCTAGCC ATCCCCCTACG AAGAAGCGGT GGCCT 35

## 25 (2) INFORMATION FOR SEQ ID NO: 14:

## (i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 1686 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## 35 (ix) FEATURE:

- (A) NAME/KEY: FY4  
 (B) LOCATION: 1...1683

## 40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

45 ATG CTG GAA CGT CTG GAA TTC GGC AGC CTC CTC CAC GAG TTC GGC CTC  
 Met Leu Glu Arg Leu Glu Phe Gly Ser Leu Leu His Glu Phe Gly Leu  
 1 5 10 15

48

50 CTG GAG GCC CCC GCC CCC CTG GAG GAG GCC CCC TGG CCC CCG CCG GAA  
 Leu Glu Ala Pro Ala Pro Leu Glu Glu Ala Pro Trp Pro Pro Pro Glu  
 20 25 30

96

GGG GCC TTC GTG GGC TTC GTC CTC TCC CGC CCC GAG CCC ATG TGG GCG  
 Gly Ala Phe Val Gly Phe Val Leu Ser Arg Pro Glu Pro Met Trp Ala  
 35 40 45

144

GAG CTT AAA GCC CTG GCC GCC TGC AGG GAC GGC CGG GTG CAC CGG GCA Glu Leu Lys Ala Leu Ala Ala Cys Arg Asp Gly Arg Val His Arg Ala 50 55 60	192
GCA GAC CCC TTG GCG GGG CTA AAG GAC CTC AAG GAG GTC CGG GGC CTC Ala Asp Pro Leu Ala Gly Leu Lys Asp Leu Lys Glu Val Arg Gly Leu 65 70 75 80	240
CTC GCC AAG GAC CTC GCC GTC TTG GCC TCG AGG GAG GGG CTA GAC CTC Leu Ala Lys Asp Leu Ala Val Leu Ala Ser Arg Glu Gly Leu Asp Leu 10 85 90 95	288
GTG CCC GGG GAC GAC CCC ATG CTC CTC GCC TAC CTC CTG GAC CCC TCC Val Pro Gly Asp Asp Pro Met Leu Leu Ala Tyr Leu Leu Asp Pro Ser 15 100 105 110	336
AAC ACC ACC CCC GAG GGG GTG GCG CGG CGC TAC GGG GGG GAG TGG ACG Asn Thr Thr Pro Glu Gly Val Ala Arg Arg Tyr Gly Glu Trp Thr 20 115 120 125	384
GAG GAC GCC GCC CAC CGG GCC CTC CTC TCG GAG AGG CTC CAT CGG AAC Glu Asp Ala Ala His Arg Ala Leu Leu Ser Glu Arg Leu His Arg Asn 25 130 135 140	432
CTC CTT AAG CGC CTC GAG GGG GAG GAG AAG CTC CTT TGG CTC TAC CAC Leu Leu Lys Arg Leu Glu Gly Glu Lys Leu Leu Trp Leu Tyr His 145 150 155 160	480
GAG GTG GAA AAG CCC CTC TCC CGG GTC CTG GCC CAC ATG GAG GCC ACC Glu Val Glu Lys Pro Leu Ser Arg Val Leu Ala His Met Glu Ala Thr 30 165 170 175	528
GGG GTA CGG CTG GAC GTG GCC TAC CTT CAG GCC CTT TCC CTG GAG CTT Gly Val Arg Leu Asp Val Ala Tyr Leu Gln Ala Leu Ser Leu Glu Leu 35 180 185 190	576
GCG GAG GAG ATC CGC CGC CTC GAG GAG GAG GTC TTC CGC TTG GCG GGC Ala Glu Glu Ile Arg Arg Leu Glu Glu Glu Val Phe Arg Leu Ala Gly 40 195 200 205	624
CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AGG GTG CTC TTT His Pro Phe Asn Leu Asn Ser Arg Asp Gln Leu Glu Arg Val Leu Phe 45 210 215 220	672
GAC GAG CTT AGG CTT CCC GCC TTG GGG AAG ACG CAA AAG ACA GGC AAG Asp Glu Leu Arg Leu Pro Ala Leu Gly Lys Thr Gln Lys Thr Gly Lys 50 225 230 235 240	720
CGC TCC ACC AGC GCC GCG GTG CTG GAG GCC CTA CGG GAG GCC CAC CCC Arg Ser Thr Ser Ala Ala Val Leu Glu Ala Leu Arg Glu Ala His Pro 55 245 250 255	768
ATC GTG GAG AAG ATC CTC CAG CAC CGG GAG CTC ACC AAG CTC AAG AAC	816

	Ile Val Glu Lys Ile Leu Gln His Arg Glu Leu Thr Lys Leu Lys Asn		
	260	265	270
5	ACC TAC GTG GAC CCC CTC CCA AGC CTC GTC CAC CCG AGG ACG GGC CGC		864
	Thr Tyr Val Asp Pro Leu Pro Ser Leu Val His Pro Arg Thr Gly Arg		
	275	280	285
10	CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GGG AGG CTT AGT		912
	Leu His Thr Arg Phe Asn Gln Thr Ala Thr Gly Arg Leu Ser		
	290	295	300
15	AGC TCC GAC CCC AAC CTG CAG AAC ATC CCC GTC CGC ACC CCC TTG GGC		960
	Ser Ser Asp Pro Asn Leu Gln Asn Ile Pro Val Arg Thr Pro Leu Gly		
	305	310	315
20	CAG AGG ATC CGC CGG GCC TTC GTG GCC GAG GCG GGT TGG GCG TTG GTG		1008
	Gln Arg Ile Arg Arg Ala Phe Val Ala Glu Ala Gly Trp Ala Leu Val		
	325	330	335
25	GCC CTG GAC TAT AGC CAG ATA GAG CTC CGC GTC CTC GCC CAC CTC TCC		1056
	Ala Leu Asp Tyr Ser Gln Ile Glu Leu Arg Val Leu Ala His Leu Ser		
	340	345	350
30	GGG GAC GAA AAC CTG ATC AGG GTC TTC CAG GAG GGG AAG GAC ATC CAC		1104
	Gly Asp Glu Asn Leu Ile Arg Val Phe Gln Glu Gly Lys Asp Ile His		
	355	360	365
35	ACC CAG ACC GCA AGC TGG ATG TTC GGC GTC CCC CCG GAG GCC GTG GAC		1152
	Thr Gln Thr Ala Ser Trp Met Phe Gly Val Pro Pro Glu Ala Val Asp		
	370	375	380
40	CCC CTG ATG CGC CGG GCG GCC AAG ACG GTG AAC TAC GGC GTC CTC TAC		1200
	Pro Leu Met Arg Arg Ala Ala Lys Thr Val Asn Tyr Gly Val Leu Tyr		
	385	390	395
45	GGC ATG TCC GCC CAT AGG CTC TCC CAG GAG CTA GCC ATC CCC TAC GAA		1248
	Gly Met Ser Ala His Arg Leu Ser Gln Glu Leu Ala Ile Pro Tyr Glu		
	405	410	415
50	GAA GCG GTG GCC TTT ATA GAG CGC TAC TTC CAA AGC TTC CCC AAG GTG		1296
	Glu Ala Val Ala Phe Ile Glu Arg Tyr Phe Gln Ser Phe Pro Lys Val		
	420	425	430
55	CGG GCC TGG ATA GAA AAG ACC CTG GAG GAG GGG AGG AAG CGG GGC TAC		1344
	Arg Ala Trp Ile Glu Lys Thr Leu Glu Gly Arg Lys Arg Gly Tyr		
	435	440	445
60	GTG GAA ACC CTC TTC GGA AGA AGG CGC TAC GTG CCC GAC CTC AAC GCC		1392
	Val Glu Thr Leu Phe Gly Arg Arg Tyr Val Pro Asp Leu Asn Ala		
	450	455	460
65	CGG GTG AAG AGC GTC AGG GAG GCC GCG GAG CGC ATG GCC TTC AAC ATG		1440
	Arg Val Lys Ser Val Arg Glu Ala Ala Glu Arg Met Ala Phe Asn Met		

	465	470	475	480	
5	CCC GTC CAG GGC ACC GCC GCC GAC CTC ATG AAG CTC GCC ATG GTG AAG Pro Val Gln Gly Thr Ala Ala Asp Leu Met Lys Leu Ala Met Val Lys 485		490		1488
10	CTC TTC CCC CGC CTC CGG GAG ATG GGG GCC CGC ATG CTC CTC CAG GTC Leu Phe Pro Arg Leu Arg Glu Met Gly Ala Arg Met Leu Leu Gln Val 500	505		510	1536
15	CAC GAC GAG CTC CTC CTG GAG GCC CCC CAA GCG CGG GCC GAG GAG GTG His Asp Glu Leu Leu Glu Ala Pro Gln Ala Arg Ala Glu Glu Val 515	520		525	1584
20	GCG GCT TTG GCC AAG GAG GCC ATG GAG AAG GCC TAT CCC CTC GCC GTG Ala Ala Leu Ala Lys Glu Ala Met Glu Lys Ala Tyr Pro Leu Ala Val 530	535		540	1632
25	CCC CTG GAG GTG GAG GTG GGG ATG GGG GAG GAC TGG CTT TCC GCC AAG Pro Leu Glu Val Glu Val Gly Met Gly Glu Asp Trp Leu Ser Ala Lys 545	550	555	560	1680
30	GGT TAG Gly *				1686

## Claims

- 35 1. An enzymatically active DNA polymerase having between 540 and 582 amino acids having a tyrosine at a position equivalent to position 667 of Taq DNA polymerase, wherein said polymerase lacks 5' to 3' exonuclease activity, and wherein said polymerase has at least 95% homology in its amino acid sequence to the DNA polymerase of Thermus aquaticus, Thermus flavus or Thermus thermophilus, and wherein said polymerase forms a single polypeptide band or an SDS polyacrylamide gel.
- 40 2. The polymerase of claim 1 wherein the amino acid sequence of said polymerase includes less than 3 conservative amino acid changes compared to one said DNA polymerase of said named Thermus species.
- 45 3. The polymerase of claim 1 wherein the amino acid sequence of said polymerase includes less than 3 additional amino acids compared to one said DNA polymerase of said named Thermus species at its N-terminus.
4. The polymerase of claim 1 selected from the group consisting of FY2, FY3 and FY4.
5. Purified nucleic acid encoding the DNA polymerase of any of claims 1-4.
- 50 6. Method for sequencing DNA comprising the step of generating chain terminated fragments from the DNA template to be sequenced with a DNA polymerase of any of claims 1-4 in the presence of at least one chain terminating agent and one or more nucleotide triphosphates, and determining the sequence of said DNA from the sizes of said fragments.
- 55 7. Kit for sequencing DNA comprising a DNA polymerase of any of claims 1-4 and a pyrophosphatase.
8. The kit of claim 7 wherein said pyrophosphatase is thermostable.

9. Apparatus for DNA sequencing having a reactor comprising a DNA polymerase of any of claims 1-4 and a band separator.

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FIG. 1  
(sheet 1)

DNA sequence 1686 b.p. acgctggagagg ... gccaaggagtga linear

1/1 31/11  
 atg ctg gag agg ctt gag ttt ggc agc ctc ccc cac gag ttc ggc ctt ctg gaa agc ccc  
 M L E R L E F G S L L H E F G L L E S P  
 61/21 91/31  
 aag gcc ctg gag gag gca ccc tgg ccc ccg ccc gaa ggg gcc ttc gca ggc ttt gtc ctt  
 K A L E E A P W P P P E G A F V G F V L  
 121/41 151/51  
 ccc cgc aag gag ccc atg tgg gcc gac ctt ctg gcc gcc gcc agg ggg ggc cgg  
 S R K E P M W A D L L A L A A A R G G R  
 181/61 211/71  
 gcc cac cgg gcc ccc gag ccc tat aaa gcc ccc agg gac ctg aag gag gca ggg ggg ccc  
 V H R A P E P Y K A L R D L K E A R G L  
 241/81 271/91  
 ccc gcc aaa gac ctg agc gtc ctg gcc ctg agg gac ggc ctt ggc ccc ccc ggc gac  
 L A K D L S V L A L R E G L G L P P G D  
 301/101 331/111  
 gac ccc atg ctc ctc gcc tac ctc ctg gac ccc aac acc acc ccc gag ggg gtc gcc  
 D P M L L A Y L L D P S N T T P E G V A  
 361/121 391/131  
 cgg cgc tac ggc ggg gag tgg acg gag gac ggg gtc gcc ccc ccc ggc gac  
 R Y G C E W T E E A G E R A A L S E R  
 421/141 451/151  
 ccc ttc gcc aac ctg tgg ggg agg ctt gag ggg gag gag agg ccc ccc tgg ccc tac cgg  
 L F A N L W G R L E G E E R L L W L Y R  
 481/161 511/171  
 gag gtg gag agg ccc ctc tcc get gtc ctg gcc cac acg gag gcc acg ggg gtc ccc  
 E V E R P L S A V L A H M E A T G V R L  
 541/181 571/191  
 gac gtg gcc tac ctc agg gcc ttc ctg gag gtg gcc gag gag acc gcc ccc ccc gag  
 D V A Y L R A L S L E V A E E I A R L E  
 601/201 631/211  
 gcc gag gtc ttc cgc ctg gcc cac ccc ctc aac acc ccc aac tcc cgg gac cag ccc gaa  
 A E V F R L A G H P F N L N S R D Q L E  
 661/221 691/231  
 agg gcc ccc ttt gac gag cta ggg ctt ccc gcc acc ggc aag acg gag aag acc ggc aag  
 R V L F D E L G L P A I G K T E K T G K  
 721/241 751/251  
 cgc tcc acc aac ggc gcc gtc ctg gag gcc ccc ccc acc gtc gag aag  
 R S T S A A V L E A L R E A H P I V E K  
 781/261 811/271  
 atc ctg cag tac cgg gag ctc acc aag ctg aag agc acc tac att gac ccc ttc ccc gac  
 I L Q Y R E L T K L K S T Y I D P L P D  
 841/281 871/291  
 ccc acc cac ccc agg acg ggc cgc ccc cac acc cgc ccc aac cag acg gcc acg gcc acg  
 L I H P R T G R L H T R F N Q T A T A T  
 901/301 931/311  
 ggc agg cta agt aac tcc gat ccc aac ctc cag aac atc ccc gcc cgc acc ccc ctt ggg  
 G R L S S S D P N L O N I P V R T P L G  
 961/321 991/331  
 cag agg acc cgc cgg gcc ttc atc gcc gag gag ggg tgg cca ttc gca ggc gcc ccc gac tac  
 Q R I R R A F I A E E G W L L V A L D Y  
 1021/341 1051/351  
 agc cag aca gag ccc agg gtc ctg gcc cac ccc ccc ggc gac gag aac ctg acc cgg gtc  
 S Q I E L R V L A H L S G D E N L I R V  
 1081/361 1111/371  
 ttc cag gag ggg cgg gac acc cac acg gag acc gcc aac acg tgg atg ttc ggc gcc ccc cgg  
 F Q E G R D I H T E T A S W M F G V P R  
 1141/381 1171/391  
 gag gcc gtg gac ccc ctg acg cgc cgg gcg gcc aag acc acc aac tac ggg gtc ccc tac  
 E A V D P L H R A A K T I N Y G V L Y  
 1201/401 1231/411  
 ggc atg tcg gcc cac cgc ctc tcc cag gag cta gcc acc ccc tac gag ggg gtc ccc  
 C M S A H D I C C F

1261/421 1291/431  
 ttc att gag cgc tac ttt cag agc ttc ccc aag gtc cgg gcc tgg att gag aag acc ctg  
 F I E R Y F Q S F P K V R A W I E K T L  
 1321/441 1351/451  
 gag gag ggc agg agg cgg ggg tac gtc gag acc ccc ttc ggc cgc cgc cgc tac gtc cca  
 E E G R R R G Y V E T L F G R R R Y V P  
 1381/461 1411/471  
 gac cta gag gcc cgg gtc aag agc gtc cgg gag gcg gcc gag cgc atg gcc ttc aac atg  
 D L E A R V K S V R E A A E R M A F N M  
 1441/481 1471/491  
 ccc gtc cag ggc acc gcc gcc gac ctc atg aag ctc gct atg gtc aag ctc ttc ccc agg  
 P V Q G T A A D L M K L A M V K L F P R  
 1501/501 1531/511  
 ctg gag gaa atg ggg gcc agg atg ctc ctt cag gtc cac gac gag ctg gtc ctc gag gcc  
 L E E M G A R M L L Q V H D E L V L E A  
 1561/521 1591/531  
 cca aaa gag agg gcg gag gcc gtc cgg ctg gcc aag gag gtc atg gag ggg gtc tac  
 P K E R A E A V A R L A K E V M E G V Y  
 1621/541 1651/551  
 ccc ctg gcc gtc ccc ctg gag gtc gag gtc ggg ata ggg gag gac tgg ctc tcc gcc aag  
 P L A V P L E V E V G I G E D W L S A K  
 1681/561 .  
 gag tga  
 E

FIG. 1  
(sheet 2)

FIG. 2  
(sheet 1)

DNA sequence 1689 b.p. atggctctggaa ... gccaaggagtga linear

1/1 31/11  
atg gct ctg gaa cgt ctg gag ttt ggc agc ctc ccc cac gag ttc ggc ctt ctg gaa agc  
M A L E R L E F G S L B H E F G L L E S  
61/21 91/31  
ccc aag gcc ctg gag gag gec ccc tgg ccc ceg cgg gaa ggg gec ttc gtc ggc ttc gtc  
P K A L E E A P W P P P E G A F V G F V  
121/41 151/51  
ctt tcc cgc aag gag ccc atg tgg gcc gat ctt ctg gcc ctg gcc gec agg ggg ggc  
L S R K E P M W A D L L A L A A A R G G  
181/61 211/71  
cgg gtc cac cgg gcc ccc gag cct tat aaa gcc ctc agg gac ctg aag gag gcg cgg ggg  
R V H R A P E P Y K A L R D L K E A R G  
241/81 271/91  
ctt ctc gcc aaa gac ctg agc gtc ctg gcc ctg agg gaa ggc ctt ggc ctc ccc ccc ggc  
L L A K D L S V L A L R E G L G L P P G  
301/101 331/111  
gac gac ccc atg ctc ctc gcc tac ccc ctg gac cct tcc aac acc acc ccc gag ggg gtc  
D D P M L L A Y L L D P S N T T P E G V  
361/121 391/131  
gcc cgg cgc tac ggc ggg gag tgg acg gag gag gcg ggg gag cgg gcc gec ctt tcc gag  
A R R Y G G E W T E E A G E R A A L S E  
421/141 451/151  
agg ctc ttc gcc aac ctg tgg ggg agg ctt gag ggg gag gag agg ctc ctc tgg ccc tac  
R L F A N L W G R L E G E E R L L W L Y  
481/161 511/171  
cgg gag gtc gag agg ccc ctt ccc gtc ctg gcc cac atg gag gec acg ggg gtc cgc  
R E V E R P L S A V L A H M E K T G V R  
541/181 571/191  
ctg gac gtc gcc tat ctc agg gec ttc ctg gag gtc gec gag gag acc gec ctc  
L D V A Y L R A L S L E V A E E I A R L  
601/201 631/211  
gag gcc gag gtc ttc cgc ctg gcc ggc cac ccc ttc aac ctc aac tcc cgg gac gag ctg  
E A E V F R L A G H P F N L N S R D Q L  
661/221 691/231  
gaa agg gtc ctc ttt gac gag cca ggg ctt ccc gec acc ggc aag acg gag aag acc ggc  
E R V L F D E L G L P A I C K T E K T G  
721/241 751/251  
aag cgc tcc acc agc gec gec gtc ctg gag gec ctc cgc gag gec ccc acc ccc gtc gag  
K R S T S A A V L E A L R E A H P I V E  
781/261 811/271  
aag atc ctg cag tac cgg gag ctc acc aag ctg aag age acc tac att gac ccc ttg cgg  
K I L Q Y R E L T K L K S T Y I D P L P  
841/281 871/291  
gac ctc atc cac ccc agg acg ggc cgc ctc cac acc cgc tcc aac cag acg gec acg gec  
D L I H P R T C R L H T R F N Q T A T A  
901/301 931/311  
acg ggc agg cta agt agc tcc gat ccc aac ctc cag aac acc ccc gtc cgc acc ccc ctt  
T G R L S S S D P N L Q N I P V R T P L  
961/321 991/331  
ggg cag agg atc cgc cgg gec ttc atc gec gag gag ggg tgg cta ctg gtc gec ccc gac  
G Q R I R R A F I A E E C W L L V A L D  
1021/341 1051/351  
tat agc cag ata gag ctc agg gtc ctg gec cac ctc tcc ggc gac gag aac ctg acc cgg  
Y S Q I E L R V L A H L S G D E N L I R  
1081/361 1111/371  
gtc ttc cag gag ggg cgg gac atc cac acg gag acc gec agc tgg atg ttc gec gec ccc  
V F Q E G R D I H T E T A S W H P G V P  
1141/381 1171/391  
cgg gag gec gtc gac ccc ctg atg cgc cgg gec gec aag acc acc aac tac ggg gec ccc  
R E A V D P L M R R A A K T I N Y G V L  
1201/401 1231/411  
cac ggc atg tcc gec cac cgc ctc tcc cag gag cta gec acc ccc tac gag gag gec cag  
Y G N S A H R L S Q E L A I P Y E E A Q

1261/421

gcc ttc att gag cgc tac ttt cag agc ttc ccc aag gtg cgg gcc tgg att gag aag acc  
 A F I E R Y F Q S F P K V R A W I E K T  
 1321/441

1351/451

atg gag gag ggc agg agg cgg ggg tac gtg gag acc ctc ttc ggc cgc cgc cgc tac gtg  
 L E E G R R R G Y V E T L F G R R R Y V

1381/461

1411/471

cca gac cta gag gcc cgg gtg aag agc gtg cgg gag ggc gag cgc atg gcc ttc aac  
 P D L E A R V K S V R E A A E R M A F N

1441/481

1471/491

atg ccc gtc cag ggc acc gcc gac ctc atg aag ctg gct atg gtg aag ctc ttc ccc  
 M P V Q G T A A D L M K L A M V K L F P

1501/501

1531/511

agg ctg gag gaa atg ggg gcc agg atg ctc ctt cag gtc cac gac gag ctg gtc ctc gag  
 R L E E M G A R M L L Q V H D E L V L E

1561/521

1591/531

gcc cca aaa gag agg gcg gag gcc gtg gcc cgg ctg gcc aag gag gtc atg gag ggg gtg  
 A P K E R A E A V A R L A K E V M E G V

1621/541

1651/551

tat ccc ctg gcc gtg ccc ctg gag gtg gag gtg ggg ata ggg gag gac tgg ctc tcc gcc  
 Y P L A V P L E V E V G I G E D W L S A

1681/561

aag gag tga  
 K E \*

FIG. 2  
 (sheet 2)

FIG. 3  
(sheet 1)

DNA sequence 2496 b.p. atggcgatgtt ... gccaaggagtag linear

1/1 31/11  
 atg gcg atg ctt ccc ctc ttt gag ccc aaa ggc cgc gtg ctc ctg gtg gac ggc cac cac  
 M A M L P L F E P K G R V L L V D G H H  
 61/21 91/31  
 ctg gcc tac cgc acc ttc ttt gcc ctc aag ggc ctc acc acc agc cgc ggc aaa ccc gtt  
 L A Y R T F F A L K G L T T S R G E P V  
 121/41 151/51  
 cag gcg gtc tac ggc ttc gcc aaa aag ctc ctc aag gcc ctg aag gag gac ggg gac gtg  
 Q A V Y G F A K S L L K A L K E D G D V  
 181/61 211/71  
 gtg gtg gtg gtc ttt gac gcc aag gcc ccc ttc ctc cgc cac gag gcc tac gag gac tac  
 V V V V F D A K A P S F R H E A Y E A Y  
 241/81 271/91  
 aag gca ggc cgg gcc ccc acc ccg gag gac ttt ccc cgg cag ctg gcc ctc atc aag gag  
 K A G R A P T P E D F P R Q L A L I K E  
 301/101 331/111  
 ttg gtg gac ctc cta ggc ctt gtg cgg ctg gag gtc ccc ggc ttt gag gca gac gac gtc  
 L V D L L G L V R L E V P G F E A D D V  
 361/121 391/131  
 ctg gcc acc ctc gcc aag cgg gca gaa aag gag ggg tac gag gtg cgc acc ctc act gcc  
 L A T L A K R A E K E G Y E V R I L T A  
 421/141 451/151  
 gac cgc gac ctc tac cag ctc ctt ctg gag cgc atc gcc atc ctc cac cct gag ggg tac  
 D R D L Y Q L L S E R I A I L H P E C Y  
 481/161 511/171  
 ctg atc acc ccg gca tgg ctt tac gag aag tac ggc ctg cgc ccg gag cag tgg gtg gac  
 L I T P A W L Y E K Y G L R P E Q W V D  
 541/181 571/191  
 tac cgg gcc ctg gca ggg gac ccc tcg gat aac atc ccc ggg gtg aag ggc acc ggg gag  
 Y R A L A G D P S D N I P G V K G I G E  
 601/201 631/211  
 aag acc gcc cag agg ctc atc cgc gag tgg ggg agc ctg gaa aac ctc ttc cag cac ctg  
 K T A Q R L I R E W G S L E N L F Q H L  
 661/221 691/231  
 gac cag gtc aag ccc tcc ttt cgg gag aag ctc cag gca ggc atg gag gcc ctg gcc ccc  
 D Q V K P S L R E K L Q A G M E A L A L  
 721/241 751/251  
 tcc cgg aag ctt ccc cag gtg cac act gac ctg ccc ctg gag gtg gac ttc ggg agg cgc  
 S R K L S Q V H T D L P L E V D F G R R  
 781/261 811/271  
 cgc aca ccc aac ctg gag ggt ctg cgg gct ttt ctg gag cgg ctg gag tcc gga agc ccc  
 R T P N L E G L R A F L E K L E F G S L  
 841/281 871/291  
 ctc cac gag ttc ggc ctc ctg gag ggg ccg aag gca gca gag gag gcc ccc tgg ccc ccc  
 L H E F G L L E G P K A A E E A P W P P  
 901/301 931/311  
 ccg gaa ggg gct ttt ttt ggc ttt tcc ttt tcc cgt ccc gag ccc atg tgg gca gag ccc  
 P E G A F L G F S F S R P E P M W A E L  
 961/321 991/331  
 ctg gcc ctg gct ggg gca ggg tgg gag ggg cgc ctc cat cgg gca caa gac ccc ccc agg ggc  
 L A L A G A W E G R L H R A Q D P L R G  
 1021/341 1051/351  
 ctg agg gac ccc aag ggg gtc cgg gga atc ctg gca aag gac ctg gca ggg gct tgg gca ctg  
 L R D L K G V R G I L A K D L A V L A L  
 1081/361 1111/371  
 cgg gag ggc ctg gac ctc ttc cca gag gac gac ccc atg ctc ctg gca tac ccc ctg gac  
 R E G L D L F P E D D P M L L A Y L L D  
 1141/381 1171/391  
 ccc tcc aac acc acc ccc gag ggg gca cgg cgt tac ggg ggg gag tgg acg gag gat  
 P S N T T P E G V A R R Y G G E W T E D  
 1201/401 1231/411  
 gca ggg gag agg gca ctc ctg gca gag gac ccc ctc cag acc ctc aag gag cgc ccc aac  
 A G E R A L L A E R L F Q T L K E R L K

1261/421  
gga gaa gaa cgc ctg ctt tgg ctt tac gag gag gtg gag aag ccg ctt tcc cgg gtg ttc  
G E E R L L W L Y E E V E K P L S R V L  
1321/441  
gcc cgg atg gag gcc acg ggg gtc egg ctg gag gac gtg gec tac ctc cag gcc ctc tcc ctg  
A R M E A T G V R L D V A Y L Q A L S L  
1381/461  
gag gtg gag gcg gag gtg cgc cag ctg gag gag gag gtc ttc cgc ctg gcc ggc eac ccc  
E V E A E V R Q L E E E V F R L A G H P  
1441/481  
ttc aac ctc aac tcc cgc gac cag ctg gag cgg gtg ctc ttt gac gag ctg ggc ctg ctc  
F N L N S R D Q L E R V L F D E L G L P  
1501/501  
gcc atc ggc aag acg gag aag acg ggg aaa cgc tcc acc agc gct gcc gtg ctg gag gce  
A I G K T E K T G K R S T S A A V L E A  
1561/521  
ctg cga gag gcc cac ccc atc gtg gac cgc atc ctg cag tac cgg gag ctc acc aag ctc  
L R E A H P I V D R I L Q Y R E L T K L  
1621/541  
aag aac acc tac ata gac ccc ctg ccc gcc ctg gtc cac ccc aag acc ggc cgg ctc cac  
K N T Y I D P L P A L V H P K T G R L H  
1681/561  
acc cgc ttc aac cag acg gcc acc gcc acg ggc agg ctt tcc agc tcc gac ccc aac ctc  
T R F N Q T A T A T G R L S S S D P N L  
1741/581  
cag aac atc ccc gtg cgc acc cct ctg ggc gag cgc atc cgc cga gcc ttc gtg gcc gag  
Q N I P V R T P L G Q R I R R A F V A E  
1801/601  
gag ggc tgg gtg ctg gtg gtc ttg gac tac agc cag acc gag ctt cgg gtc ctg gcc cac  
E G W V L V V L D Y S Q I E L R V L A H  
1861/621  
ctc tcc ggg gac gag aac ctg atc cgg gtc ttc cag gag ggg agg gac atc cac acc cag  
L S G D E N L I R V F Q E G R D I H T Q  
1921/641  
acc gcc agc tgg atg ttc ggc gtt tcc ccc gaa ggg gta gac cct ctg atc cgc cgg ggc  
T A S W M F G V S P E G V D P L M R R A  
1981/661  
gcc aag acc atc aac ttc ggg gtg ctc tac ggc atg tcc gcc cac cgc ctc tcc ggg gag  
A K T I N F G V L Y G M S A H R L S G E  
2041/681  
ctt tcc atc ccc tac gag gag ggc gtg gec ttc att gag cgc tac ttc cag agc tac ccc  
L S I P Y E E A V A F I E R Y F Q S Y P  
2101/701  
aag gtg cgg gec tgg att gag ggg acc ctc gag gag ggc cgc cgg cgg ggg tat gtg gag  
K V R A W I E G T L E E G R R G Y V E  
2161/721  
acc ctc ttc ggc cgc cgg cgc tat gtg ccc gag ctc aac gcc cgg gtg aag agc gtg cgc  
T L F G R R R Y V P D L N A R V K S V R  
2221/741  
gag ggc gcg gag cgc atg gec ttc aac atg cgg gtc gag ggc acc gcc gac ctc acg  
E A A E R M A F N M P V Q G T A A D L M  
2281/761  
aag ctg gcc atg gtg cgg ctt ttc ccc cgg ctt cag gaa ctg ggg ggc agg atg ctt ctg  
K L A M V R L F P R L Q E L G A R M L L  
2341/781  
cag gtg cac gac gag ctg gtc ctc gag gcc ccc aag gac cgg ggc gag aag gta gcc gct  
Q V H D E L V L E A P K D R A E R V A A  
2401/801  
ttg gcc aag gag gtc atg gag ggg gtc tgg ccc ctg cag gtg ccc ctg gag gtg gag gtg  
L A K E V M E G V W P L Q V P I E V E V  
2461/821  
ggc ctg ggg gag gac tgg ctc tcc gcc aag gag tag  
G L G E D W L S A K E

FIG. 3  
(sheet 2)

FIG. 4  
(sheet 1)

XNA sequence 2505 b.p. ATGGAGGCGATG ... GCCAANGGGTTAG linear

coding sequence of *T. thermophilus* DNA polymerase as submitted by D. Gelfand in WO 91/09950 PCT/US91/071

1/1  
VIG GAG GCG ATG CTT CGC CTC TTT GAA CCC AAA GGC CGG GTC CTC CTG GTG GAC GGC GGC  
1 E A M L P L F E P K G R V L L V D G H  
61/21  
91/31  
CAC CTG GCC TAC CGC ACC TTC TTC GCC CTG AAG GGC CTC ACC ACG AGC CGG GGC GAA CGC  
H L A Y R T F F A L K G L T T S R G E P  
121/41  
151/51  
CTG CAG GGG GTC TAC CGC TTC CGC AAG AGC CTC CTC AAG GGC CTG AAG GAG GAC GAC GGG TAC  
V Q A V Y G F A K S L L K A L K E D G Y  
181/61  
211/71  
AAG GCC GTC TTC CTG GTC TTT GAC GCC AAG GCC CCC TCC TTC CGC CAC GAG GGC TAC GAG  
K A V F V V F D A K A P S F R H E A Y E  
241/81  
271/91  
GCC TAC AAC CGG CGG AGG GCC CGG ACC CCC GAG GAC TTC CGC CGG CAG CTC GCC CTC ATC  
A Y K A G R A P T P E D F P R Q L A L I  
301/101  
331/111  
AAC GAG CTG CTG GAC CTC CTG CGG TTT ACC CGC CTC GAG GTC CGC CGC TAC GAG GCG GAC  
K E L V D L L G F T R L E V P G Y E A D  
361/121  
391/131  
CAC GTT CTC CGC ACC CTG CGC AAG AAG CGG GAA AAG CAG CGG TAC GAG GTG CGC ATC CTC  
D V L A T L A K K A E K E G Y E V R I L  
421/141  
451/151  
ACC GCC GAC CGC GAC CTC TAC CAA CTC CTC TCC GAC CGC CTC CGC CTC CTC CAC CGC GAC  
T A D R D L Y Q L V S D R V A V L H P E  
481/161  
511/171  
GGC CAC CTC ATC ACC CGG GAG TGG CTT TGG GAG AAG TAC GGC CTC AGG CGG GAG CGG TGG  
G H L I T P E W L W E K Y G L R P E Q W  
541/181  
571/191  
GTC GAC TTC CGC CGC CTC GTG CGG GAC CGC TCC GAC AAC CTC CGC CGG GTC AAG GCC ATC  
V D P R A L V G D P S D N L P G V K G I  
601/201  
631/211  
GGG GAG AAG ACC CGC CTC AAG CTC CTC AAC GAG TGG GGA AAG CTC GAA AAC CTC CTC AAC  
G E K T A L K L L K E W G S L E N L L K  
661/221  
691/231  
AAC CTG GAC CGG GTA AAG CCA GAA AAC GTC CGG GAG AAG ATC AAC GGC CAC CTG GAA GAC  
N L D R V K P E N V R E K I K A H L E D  
721/241  
751/251  
CTC AGG CTC TCC TTG GAG CTC TCC CGG GTG CGC ACC GAC CTC CGC CTG GAG GTG GAC CTC  
L R L S L E L S R V R T D L P L E V D L  
781/261  
811/271  
GCC CAG GGG CGG GAG CGC GAC CGG GAG CGG CTT AGG GCT TTC CTC GAG AGG CTC GAG TTC  
A Q G R E P D R E G L R A F L E R L E F  
841/281  
871/291  
GGC AGC CTC CTC CAC GAG TTC CGC CTC CTG GAG GCC CGC CCC CTC GTC CTC CTC CTC AAC TGG  
G S L L H E F G L L E A P A P L E E A P  
901/301  
931/311  
TGG CCC CGG CGG GAA CGG GGC TTC GTG GGC TTC GTC CTC TCC CGC CGC AAC GAC CTC CTC AAC TGG  
W P P P E G A F V G F V L S R P E P M W  
961/321  
991/331  
GCG GAG CTT AAA CGC CTG CGC CGC TCC AGG GAC GGC CGG GTC CAC CGG GCA GCA GAC CGC  
A E L K A L A A C R D G R V H R A A D P  
1021/341  
1051/351  
TTG CGG CGG CTA AAG GAC CTC AAC GAG GTC CGG GGC CTC CTC CGC AAC GAC CTC CTC CGC  
L A G L K D L K E V R G L L A K D L A V  
1081/361  
1111/371  
TTG GCC TCG AGG GAG CGG CTA GAC CTC GTG CCC CGG GAG CAC CGC AAC GAC CTC CTC CGC AAC TAC  
L A S R E G L D L V P G D D P M L L A Y  
1141/381  
1171/391  
CTC CTG GAC CGC CCC TCC AAC ACC ACC CGG GAG CGG CTG CGC CGS CSC TAC 1021/360 GAG TGG  
L L D P S N T T P E G V A R R Y U C E W

1201/401 1231/411  
 ACG GAG GAC CCC CCC CAC CGG GCG CTC CTC TCG GAG AAG CTC CAT CGG AAC CTC CTC AAG  
 T E D A A H R A L L S E R L H R N L L K  
 1261/421 1291/431  
 CGC CTC GAG GGG GAG GAG AAG CTC CTC TCG CTC TAC CAC GAG GTG GAA AAG CCC CTC TCC  
 R L E G E E K L L W L Y H E V E K P L S  
 1321/441 1351/451  
 CGG GTC CTG GCC CAC ATG GAG GCG ACC CGG GTA CGG CTG GAC GTG CCC TAC CTT CAG GCC  
 R V L A H M E A T G V R L D V A Y L Q A  
 1381/461 1411/471  
 CTT TCC CTG GAG CTT CGG GAG GAG ATC CGC CGC CTC GAG GAG GTC TTC CGC TTG GCG  
 L S L E L A E E I R R L E E E V, F R L A  
 1441/481 1471/491  
 GGC CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA AAG GTG CTC TTT GAC GAG CTT  
 G H P F N L N S R D Q L E R V L F D E L  
 1501/501 1531/511  
 AGG CTT CCC CCC TTG CGG AAG ACG CAA AAG ACA GGC AAG CGC TCC ACC AGC GGC GCG GTG  
 R L P A L G K T Q K T G K R S T S A A V  
 1561/521 1591/531  
 CTG GAG GCG CTA CGG GAG GCG CAC CCC ATC GTG GAG AAG ATC CTC CAG CAC CGG GAG CTC  
 L E A L R E A H P I V E K I L Q H R E L  
 1621/541 1651/551  
 ACC AAG CTC AAG AAC ACC TAC GTG GAC CCC CTC CCA AGC CTC GTC CAC CGG AGG ACG GGC  
 T K L K N T Y V D P L P S L V H P R T G  
 1681/561 1711/571  
 CGC CTC CAC ACC CGC TTC AAC CAG ACG GGC ACG GGC ACG GGG AGG CTT AGT AGC TCC GAC  
 R L H T R F N Q T A T A T G R L S S S D  
 1741/581 1771/591  
 CCC AAC CTG CAG AAC ATC CCC GTC CGC ACC CCC TTG GCG CAG AGG ATC CGG CGG CCC TTC  
 P N L Q N I P V R T P L G Q R I R R A F  
 1801/601 1831/611  
 GTG GCG GAG GCG GGT TGG GCG TTG GTG GCG CTG GAC TAT ACG CAG ATA GAG CTC CGC GTC  
 V A E A G W A L V A L D Y S Q I E L R V  
 1861/621 1891/631  
 CTC GCC CAC CTC TCC CGG GAC GAA AAC CTG ATC AGG GTC TTC CAG GAG CGG AAG GAC ATC  
 L A H L S G D E N L I R V F Q E G K D I  
 1921/641 1951/651  
 CAC ACC CAG ACC GCA AGC TGG ATG TTC CGC CTC CCC CGG CAG CGC CCC GTG CAC CCC CTG ATC  
 H T Q T A S W M F G V P P E A V D P L M  
 1981/661 2011/671  
 CGC CGG GCG CCC AAG ACG GTG AAC TTC CGC GTC CTC TAC CGC ATG TCC CGC CAT AGG CTC  
 R R A A K T V N F G V L Y G M S A H R L  
 2041/681 2071/691  
 TCC CAG GAG CTT GCC ATC CCC TAC GAG GAG GCG GTG CCC TTT ATA GAG CGC TAC TTC CAA  
 S Q E L A I P Y E E A V A F I E R Y F Q  
 2101/701 2131/711  
 AGC TIC CCC AAG GTG CGG CCC TTG ATA GAA AAG ACC CTG CAG CAG GGG AGG AAG CGG CGC  
 S F P K V R A W I E K T L E E G R K R G  
 2161/721 2191/731  
 TAC GTC GAA ACC CTC TTC CGA AGA AGG CGC TAC GTG CCC CGC CTC AAC CGC CGG GTG AAG  
 Y V E T L F G R R R Y V P D L N A R V K  
 2221/741 2251/751  
 AGC GTC AGG GAG CCC CGG CAG CGC ATG CCC ATG CTC CTC GAG CGC CGC AGC CGC CGC  
 S V R E A A E R M A F N M P V Q G T A A  
 2281/761 2311/771  
 GAC CTC ATG AAG CTC CCC ATG GTG AAG CTC TTC CCC CGC CTC CGG GAG ATG GGG CGC CGC  
 D L M K L A M V K L F P R L R E M G A R  
 2341/781 2371/791  
 ATG CTC CTC CAG GTC CAC GAC GAG CTC CTC GTG GAG CGC CGC CAA CGG CGG CGC GAG GAG  
 M L L Q V H D E L L E A P Q A R A E E  
 2401/801 2431/811  
 GTG CGG GCT TTG GCC AAG GAG GCG ATG GAG AAG CCC TAT CCC CTC CGC GTG CGC CTC GAG  
 V A A L A K E A M E K A Y P L A V P L E  
 2461/821 2491/831  
 GTG GAG GTG GGG ATG GGG CAG GAC TGG CTT TCC CGC AAG CGT TAG  
 V E V G M G E D W L S A K C

FIG. 5

(Sheet 1)

## DNA and protein sequence of the coding region of pMR8, encoding FY4

1/1	31/11
ATG CTG GAA CGT CTG GAA TTC GGC AGC CTC CTC CAC GAG TTC GGC CTC CTG GAG GCC CCC	
M L E R L E F G S L L H E F G L L E A P	
61/21	91/31
GCC CCC CTG GAG GAG GCC CCC TGG CCC CCG CCG GAA GGG GCC TTC GTG GGC TTG CTC CTC	
A P L E E A P W P P P E G A F V G F V L	
121/41	151/51
TCC CGC CCC GAG CCC ATG TGG GCG GAG CTT AAA GCC CTG GCC GCC TGC AGG GAC GGC CGG	
S R P E P M W A E L K A L A A C R D G R	
181/61	211/71
GTG CAC CGG GCA GCA GAC CCC TTG GCG GGG CTA AAG GAC CTC AAG GAG GTC CGG GGC CTC	
V H R A A D P L A G L K D L K E V R G L	
241/81	271/91
CTC GCC AAG GAC CTC GCC GTC TTG GCC TCG AGG GAG GGG CTA GAC CTC GTG CCC GGG GAC	
L A K D L A V L A S R E G L D L V P G D	
301/101	331/111
GAC CCC ATG CTC CTC GCC TAC CTC CTG GAC CCC TCC AAC ACC ACC CCC GAG GGG GTG GCG	
D P M L L A Y L L D P S N T T P E G V A	
361/121	391/131
CGG CGC TAC GGG GGG GAG TGG ACG GAG GAC GCC CAC CGG GCC CTC CTC TCG GAG AGG	
R R Y G G E W T E D A A H R A L L S E R	
421/141	451/151
CTC CAT CGG AAC CTC CTT AAG CGC CTC GAG GGG GAG AAG CTC CTT TGG CTC TAC CAC	
L H R N L L K R L E G E E K L L W L Y H	
481/161	511/171
GAG GTG GAA AAG CCC CTC TCC CGG GTC CTG GCC CAC ATG GAG GCC ACC GGG GTA CGG CTG	
E V E K P L S R V L A H M E A T G V R L	
541/181	571/191
GAC GTG GCC TAC CTT CAG GCC CTT TCC CTG GAG CTT GCG GAG GAG ATC CGC CGC CTC GAG	
D V A Y L Q A L S L E L A E E I R R L E	
601/201	631/211
GAG GAG GTC TTC CGC TTG GCG GGC CAC CCC TTC AAC CTC AAC TCC CGG GAC CAG CTG GAA	
E E V F R L A G H P F N L N S R D Q L E	
661/221	691/231
AGG GTG CTC TTT GAC GAG CTT AGG CTT CCC GCC TTG GGG AAG ACG CAA AAG ACA GGC AAG	
R V L F D E L R L P A L G K T Q K T G K	
721/241	751/251
CGC TCC ACC AGC GCC GCG GTG CTG GAG GCC CTA CGG GAG GGC CAC CCC ATC GTG GAG AAG	
R S T S A A V L E A L R E A H P I V E K	
781/261	811/271
ATC CTC CAG CAC CGG GAG CTC ACC AAG CTC AAC ACC TAC GTG GAC CCC CTC CCA AGC	
I L Q H R E L T K L K N T Y V D P L P S	
841/281	871/291
CTC GTC CAC CCG AGG ACG GGC CGC CTC CAC ACC CGC TTC AAC CAG ACG GCC ACG GCC ACG	
L V H P R T G R L H T R F N Q T A T A T	
901/301	931/311
GGG AGG CTT AGT AGC TCC GAC CCC AAC CTG CAG AAC ATC CCC GTC CGC ACC CCC TTG GGC	
G R L S S S D P N L Q N I P V R T P L G	
961/321	991/331
CAG AGG ATC CGC CGG GCC TTC GTG GCC GAG GCG GGT TGG GCG TTG GTG GCC CTG GAC TAT	
Q R I R R A F V A E A G W A L V A L D Y	
1021/341	1051/351

FIG 5.  
(Sheet 2)

AGC CAG ATA GAG CTC CGC GTC CTC GCC CAC CTC TCC GGG GAC GAA AAC CTG ATC AGG GTC  
 S Q I E L R V L A H L S G D E N L I R V  
 1081/361 1111/371  
 TTC CAG GAG GGG AAG GAC ATC CAC ACC CAG ACC GCA AGC TGG ATG TTC GGC GTC CCC CCG  
 F Q E G K D I H T Q T A S W M F G V P P  
 1141/381 1171/391  
 GAG GCC GTG GAC CCC CTG ATG CGC CGG GCG GCC AAG ACG GTG AAC TAC GGC GTC CTC TAC  
 E A V D P L M R R A A K T V N Y G V L Y  
 1201/401 1231/411  
 GGC ATG TCC GCC CAT AGG CTC TCC CAG GAG CTA GCC ATC CCC TAC GAA GAA GCG GTG GCC  
 G M S A H R L S Q E L A I P Y E E A V A  
 1261/421 1291/431  
 TTT ATA GAG CGC TAC TTC CAA AGC TTC CCC AAG GTG CGG GCC TGG ATA GAA AAG ACC CTG  
 F I E R Y F Q S F P K V R A W I E K T L  
 1321/441 1351/451  
 GAG GAG GGG AGG AAG CGG GGC TAC GTG GAA ACC CTC TTC GGA AGA AGG CGC TAC GTG CCC  
 E E G R K R G Y V E T L F G R R R Y V P  
 1381/461 1411/471  
 GAC CTC AAC GCC CGG GTG AAG AGC GTC AGG GAG GCC GCG GAG CGC ATG GCC TTC AAC ATG  
 D L N A R V K S V R E A A E R M A F N M  
 1441/481 1471/491  
 CCC GTC CAG GGC ACC GCC GGC GAC CTC ATG AAG CTC GCC ATG GTG AAG CTC TTC CCC CGC  
 P V Q G T A A D L M K L A M V K L F P R  
 1501/501 1531/511  
 CTC CGG GAG ATG GGG GCC CGC ATG CTC CTC CAG GTC CAC GAC GAG CTC CTC CTG GAG GCC  
 L R E M G A R M L L Q V H D E L L L E A  
 1561/521 1591/531  
 CCC CAA CGC CGG GCC GAG GAG GTG GCG CCT TTG GCC AAG GAG GCC ATG GAG AAG GCC TAT  
 P Q A R A E E V A A L A K E A M E K A Y  
 1621/541 1651/551  
 CCC CTC GCC GTG CCC CTG GAG GTG GAG GTG GGG ATG GGG GAG GAC TGG CTT TCC GCC AAG  
 P L A V P L E V E V G M G E D W L S A K  
 1681/561  
 GGT TAG  
 G \*



European Patent  
Office

## **EUROPEAN SEARCH REPORT**

*Application Number*

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.)
D,A	WO-A-92 06188 (BARNES WAYNE M) * the whole document *	1-9	C12N15/54 C12N9/12 C12Q1/68
A	WO-A-91 09944 (CETUS CORP) * the whole document *	1-9	
A	WO-A-94 05797 (KISELEV VSEVOLOD ;SEVERIN EVGENII (RU); KORPELA TIMO (FI)) * the whole document *	1-9	
D,A	EUR. J. BIOCHEM. (1992), 209(1), 351-5 CODEN: EJBCAI;ISSN: 0014-2956, 1992, XP000578012 RICHTER, OLIVER MATTHIAS H. ET AL: "Cloning and sequencing of the gene for the cytoplasmic inorganic pyrophosphatase from the thermoacidophilic archaeabacterium Thermoplasma acidophilum" * the whole document *	1-9	
A	WO-A-90 12111 (HARVARD COLLEGE) * the whole document *	1-9	TECHNICAL FIELDS SEARCHED (Int.Cl.)
P,X, D	EP-A-0 655 506 (PRESIDENT AND FELLOWS OF HARVARD COLLEGE, USA) * page 6, line 15 - line 17; claims 1-52 *	1,6,9	
P,X	NATURE, vol. 376, 31 August 1995, MACMILLAN JOURNALS LTD., LONDON,UK. pages 796-797, XP002009831 M.A. REEVE AND C.W. FULLER: "A novel thermostable polymerase for DNA sequencing" * the whole document *	1-9  -/-	C12N C12Q
The present search report has been drawn up for all claims			
Place of search	Date of completion of the search	Examiner	
THE HAGUE	20 September 1996	Hornig, H	
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone	I : theory or principle underlying the invention		
Y : particularly relevant if combined with another document of the same category	E : earlier patent document, but published on, or after the filing date		
A : technological background	R : document cited in the application		
O : non-patent disclosure	D : document cited for other reasons		
P : intermediate document	M : number of the same patent family, corresponding document		



European Patent  
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## EUROPEAN SEARCH REPORT

Application Number  
EP 96 30 3880

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.)
P,X	<p>AMERSHAM LIFE SCIENCE, EDITORIAL COMMENTS, vol. 22, no. 2, July 1995, pages 29-36, XP002009832</p> <p>S.B. SAMOLS ET AL.: "Thermo Sequenase; a new thermostable DNA polymerase for DNA sequencing" * the whole document *</p> <p>-----</p> <p>PROC. NATL. ACAD. SCI. U. S. A. (1995), 92(14), 6339-43 CODEN: PNASA6; ISSN: 0027-8424, 3 July 1995, XP002009833</p> <p>TABOR, STANLEY ET AL: "A single residue in DNA polymerases of the Escherichia coli DNA polymerase I family is critical for distinguishing between deoxy- and dideoxyribonucleotides" * the whole document *</p> <p>-----</p>	1-9	
P,A		1-9	TECHNICAL FIELDS SEARCHED (Int.Cl.)
<p>The present search report has been drawn up for all claims</p>			
Place of search	Date of completion of the search	Examiner	
THE HAGUE	20 September 1996	Hornig, H	
CATEGORY OF CITED DOCUMENTS		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons A : member of the same patent family, corresponding document	
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document			